

# NGS Data Analysis and Galaxy

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# This Week

Monday	Welcome, Project Intro, Basic Galaxy Usage NGS QualityControl
Tuesday	RNA-Seq - Mapping and Transcript Prediction RNA-Seq: Differential expression and Alternative Pipelines; SNP & Variant Analysis
Wednesday	<b>SNP &amp; Variant Analysis</b> <b>Chip-Seq Analysis</b>
Thursday	Genome Assembly Install your own Galaxy on Amazon Cloud
Friday	Customizing Galaxy, Galaxy Tool Shed, and Wrapping Tools for Galaxy

# Wednesday Agenda

- 8:30 **Welcome and Questions**
- 8:45 **SNP and Variant Analysis, Part III**
- 10:30 **Break**
- 11:00 **SNP and Variant Analysis, Part IV**
- 12:30 **Lunch**
- 13:30 **ChIP-Seq I**
- 15:00 **Break**
- 15:30 **ChIP-Seq II**
- 16:30 **Done**

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# ChIP-Seq: FASTQ data and quality control

<http://scriptogr.am/ohofmann>

By Shannan Ho Sui

Create a new history

Shared Data → Data Libraries

→ **ChIP-Seq Datasets**

Select everything in the **Filtered Reads** folder

Also grab **genes\_chr12.gtf** from

**UC Davis RNA-Seq Human library**

# ChIP-Seq Exercise: Mapping with Bowtie

Using Bowtie directly (could also use BWA)

NGS Mapping: → Map with Bowtie for Illumina

FASTQ file → H1hesc Nanog Rep1 chr12 qualityfiltered

Single End, hg19\_chr12

Full Parameter List

Suppress all alignments for a read if ... → 1

# ChIP-Seq Analysis: **remove unmapped reads**

**SAM Tools** → **Filter SAM**

- Click **Add a new Flag**
- Set **Type** to **The read is unmapped**
- Set flag **state** to **No.**

# ChIP-Seq Analysis: Put mapped reads in BAM

- **SAM Tools** → **SAM-to-BAM**
- Also get the the control (already mapped for us)
  - **Shared Data** → **Data Libraries** → **Aligned** →  
Import **H1hesc Input Rep1 Chr12 Mapped** into  
current history



# ChIP-Seq Analysis: Find Peaks

NGS: Peak Calling → MACS

Experiment name → MACS NanogRep1

Tag File → Nanog Rep1 BAM file

Control File → Input Rep1 BAM file

Tag Size → 36

Save shifted raw tag count → ... Save (leave res. at 10)

Check Perform the new peak detection method (future dir)

# ChIP-Seq Analysis: Visualize Results

Look at the HTML report dataset

Launch a Trackster visualization and bring in

the called peaks

the Treatment WIG

the Control WIG

the gene definitions

# ChIP-Seq Analysis: Replicates

Shared Data → Data Libraries → ChIP-Seq Datasets →  
MACS Outputs

Import **Peaks** files for

Nanog Rep 2

Pou5f1 Rep 1

Pou5f1 Rep 2

# ChIP-Seq Analysis: Unify Replicates

Operate on Genomic Intervals → Concatenate

Concatenate Nanog Rep 1 and 2

Operate on Genomic Intervals → Cluster

Use default parameters

Rename the output dataset

Finally, add the **Nanog Rep 2 Peaks**, and the **cluster** output to your visualization

# ChIP-Seq Analysis: Unify Replicates

Repeat for **Pou5f1** replicates

**Operate on Genomic Intervals** → **Concatenate**

**Concatenate Pou5f1 Rep 1 and 2**

**Operate on Genomic Intervals** → **Cluster**

Use default parameters

Rename the output dataset

Finally, add the **Pou5f1 Rep 1 & 2 Peaks**, and the **cluster** output to your visualization

# ChIP-Seq Analysis: Differential binding

Operate on Genomic Intervals → Subtract

First dataset clustered → Pou5f1

Second dataset clustered → Nanog

Return → Intervals with no overlap

# ChIP-Seq Mapping With MACS

## Further reading & Resources

[ChIP-Seq: FASTQ data and quality control](#)

by Shannan Ho Sui

[HAIB TFBS ENCODE collection](#)

[MACS Documentation](#)

[Model-based analysis of ChIP-Seq \(MACS\)](#)

by Zhang *et al.*

[Cistrome](#) and [Nebula](#) Galaxy Servers

[Nebula Tutorial](#)

by Valentina Boeva

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**Thanks**



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