The Viral Cloud Resource

Ravi Sanka, Ntino Krampis, Alex Richter, Andrey Tovchigrechko

@ GCC 2014, Baltimore





- Objective: To produce an interactive and informative display of an annotated viral genome based on provided sequences, both reads and established references.
- Inputs
 - Read sequences that can come from a variety of platforms.
 - A viral database chosen from a provided list of organisms.





- Outputs:
 - Assembled genome of the viral database's organism.
 - Standard output of the JCVI VIGOR tool.
 - An interactive JBROWSE visualization of the annotated genome.
- Consists of two stages (Galaxy tools)
 - Viral Assembly
 - JCVI VIGOR



- What is it?
 - A python script that takes in provided read sequences, established reference database file of the target viral genome, and produces a FASTA file of the viral genome modified by the read sequences.
 - The means of achieving this goal were determined by the JCVI Viral Group.
- The program achieves this through the following steps.



- Step 1 Acquire Inputs
 - Input read sequences are retrieved via option parsing. User can provide and combination of 454 (sff), Illumina (fastq), Sanger (fasta and qual), and IonTorrent (fastq).
 - The name of the chosen reference is also retrieved via option parsing. The name points the script to the appropriate, preset location of the BLAST-able database.
 - Read sequences are converted to FASTQ, offset-33 format.



- Step 2 Choose Appropriate Reference Sequence
 - Reduce the read count of each input read set (one for each platform) to the number of reads needed to cover 50% of the target viral genome (this figure is stored) via read lengths.
 - This is done to decrease runtime of the rest of this step.
 - Denovo-assemble the entire set of reads (after read count reduction) via CAP3.
 - Align the contigs to the chosen reference database, which contains several genome sequences of the target virus, and select the top hit with BLASTN.

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This is the reference sequence.

- Step 3 Reference Mapping and SNP Gathering.
 - Return the original read sets (full read counts).
 - 1 or 2 of the read sets are chosen for this step, based on their sequence platforms.
 - Which read sets are chosen follow this hierarchy, from first-choice to last:
 - 454 and Illumina
 - 454 and lontorrent
 - 454 only

1.

2.

3.

4.

5.

6.

7.

- Illumina and Iontorrent
- Illumina only
- lontorrent only
 - Sanger only

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- For each read set, align to the reference sequence via BWA and gather all SNPs found via SAMTOOLS, BCFTOOLS, and VCFUTILS.
- Extract common SNPs found in both alignments via sdiff.
 - If only one read set was chosen (hierarchy choice 3, 5, 6, or 7), all SNPs are kept.



- Step 4 Producing Final Viral Genome Sequence.
 - Modify the reference sequence with the SNPs found in Step 3 via delta2seq.
 - Align each read set to the modified reference via BWA.
 - Merge the sorted BAM of each BWA execution into one BAM via SAMTOOLS.
 - Extract the consensus sequence of the total alignment via SAMTOOLS, BCFTOOLS, and VCFUTILS.
 - This consensus sequence is the final, modified viral genome sequence.



- What is it?
 - VIral Genome Orf Reader (VIGOR).
 - A gene predictor program for small viral genomes.
 - VIGOR uses a similarity-based approach to detect ORFs by similarity searches against custom reference protein sequence databases.
 - Takes into account differences between the genomic structures of viral taxonomic groups.
 - Identifies frame-shifts, ribosomal slippage, RNA editing, stop codon read-through, overlapping genes, embedded genes, and mature peptide cleavage sites.
 - Genotyping capability for influenza and rotavirus is built into the program.

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- VIGOR can be run from command-line via its Perl script wrapper.
- VIGOR is also publicly available as a webtool on JCVI's website.
- More information can be found here:
 - JCVI main VIGOR page and webtool: http://www.jcvi.org/vigor/index.php
 - Publication: http://www.biomedcentral.com/1471-2105/11/451



- For the VCR Pipeline, VIGOR is installed on the server and added as a Galaxy tool.
- The Galaxy tool has one input, a FASTA file of the sequence(s) the user wants annotated.
 - This is typically the output of Viral Assembly, but since VIGOR is a separate tool, the user can choose sequences from an outside source.



- Upon execution, the Galaxy tool executes the locally installed VIGOR on the FASTA file.
- Bundled with the VIGOR program itself, and installed alongside VIGOR during pipeline installation, are all the BLAST databases that VIGOR uses when annotating the input FASTA file.



- The Galaxy tool provides the following output files of VIGOR as output datasets:
 - ALN: CLUSTAL-W alignment file of genes found
 - CDS: FASTA file of features' nucleotide sequences
 - FS: logging data; this file is typically empty
 - PEP: FASTA file of features' peptide sequences
 - RPT: report of parameters used in run
 - TBL: tab-delimited file of discovered features
 - STATS: tab-delimited file coord/cov/ID data of features
- The Galaxy tool also uses VICVB (Venter Institute Cloud Viral Browser), a visualization module installed with VIGOR during pipeline installation, to produce an interactive JBrowser une

vcr.py

- Stands for Viral Cloud Resource.
- A python Fabric script for the following tasks:
 - Enables the environment to run the VCR pipeline.
 - Downloads and installs necessary dependencies (reference database files, 3rd-party tools, etc).
 - Installs the pipeline itself.
 - Adds the front-end files (XML and python) to the Galaxy instance (which must be present).
 - Installs the pipelines scripts on the box, and makes them accessible to the Galaxy front-end python scripts.



vcr.py has no main.

- Like a library, it consists of functions that are used by a higher-level script (fabfiles.py) as appropriate.
- A YAML config file (custom.yaml) allows outside scripts access to only a select few of these functions, which in turn make use of the vcr.py's other, deeper functions to carry out their tasks.
 - This allows the higher-level script to complete the entire installation of the pipeline via a few functions calls.
 - Keeps code compartmentalized and minimizes the number of locations to edit during modifications/updates.





Installation Methods

- install_galaxy_vcr
 - Installs the VCR Pipeline tools to the target Galaxy install.
 Downloads and properly places tools' XML/Python scripts and sets the necessary config files.
- install_viralassembly
 - Installs the scripts and dependencies of Viral Assembly on the target VM.
- install_viralvigor
 - Installs the scripts and dependencies of VIGOR on the target VM.

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- Install_vicvb
 - Installs the VICVB package.

Task Methods (install_viralassembly)

- _initialize_area_viral
 - Defines paths and environment variables for Viral Assembly.
- _add_tools_viral
 - Downloads and installs main script and dependencies of Viral Assembly in the locations set by _initialize_area_viral.
 - Also installs the bio-linux, allowing apt-get to access bioinformatic tools.
- _add_refs

Downloads and installs the BLAST-able database files of the viral reference sequences in the locations set by _initialize_area_viral.



Task Methods (install_viralvigor)

- _initialize_area_vigor
 - Defines paths and environment variables for VIGOR.
- _initialize_host
 - Defines (and creates if not present) a location for VIGOR's byproduct files.
- _add_vigor

Downloads and installs the VIGOR package in the locations set by _initialize_area_vigor.

_add_tools_vigor

Downloads and installs the required programs and libraries, not included in the initial VIGOR package, in the locations set by __initialize_area_vigor.



Common Methods

- _add_package
 - Downloads target package via wget to appropriate location and decompresses with corresponding program (tar, bz2, gzip, etc).
 - Used to download dependencies stored in Amazon S3 and FTP repositories.
- _apt_get_install
 - Executes apt-get to install target tool (i.e. csh, gawk, bwa, samtools, etc)

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- vcr.py dependencies:
 - The target box must have Galaxy installed.
 - vcr.py assumes this is "/mnt/galaxyTools/galaxy-central", but this can be changed by modifying the global variable galaxy_central.
 - Must have git, apt-get, and wget enabled.
 - This are used to install and download the needed programs and files from the various repositories.



Thanks too:

- The rest of the Team:
 - Ntino Krampis
 - Enis Afgan
 - Brad Chapman
 - Alex Richter
 - Andrey Tovchigrechko

JCVI Viral Group

- Tim Stockwell
- Danny Katzel
- Jeff Hoover

