# Introduction to Galaxy

The University of Edinburgh Edinburgh, United Kingdom 12 May 2014

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http://galaxyproject.org/





Wellcome Trust Centre for Cell Biology





**Edinburgh Bioinformatics** 







# The Agenda

- 9:00 Welcome, Galaxy Platforms
- 9:20 Basic Analysis with Galaxy
- 10:30 Break
- 10:45 RNA-Seq Example
- 12:40 Project and Community Overview

13:00 Done

# The Agenda

Goal is to demonstrate how Galaxy can help you explore and learn options, perform analysis, and then share, repeat, and reproduce your analyses.

# Not The Agenda

### This workshop will not cover

- details of how tools are implemented, or
- new algorithm designs, or
- which assembler or mapper or peak caller or ... is best for you.

While this workshop does cover RNA-Seq, we are only using that specific example to learn general principles.

### What is Galaxy?

A data integration and analysis platform A free (for everyone) web server Open source software <u>These options result in several ways to use Galaxy</u>

http://galaxyproject.org

### Galaxy is available ...

As a free (for everyone) web server integrating a wealth of tools, compute resources, terabytes of reference data and permanent storage

http://usegalaxy.org

However, a centralized solution cannot support the different analysis needs of the entire world.



Leveraging the national cyberinfrastructure for biomedical research LeDuc, et al. J Am Med Inform Assoc doi:10.1136/amiajnl-2013-002059

### Galaxy is available ...

 As a free (for everyone) web service http://usegalaxy.org

• As open source software

http://getgalaxy.org

It is installed in locations around the world

### Galaxy is available on the Cloud

CloudMan

### We are using this today. Start with a fully configured & populated (tools and data) Galaxy instance.

Allows you to scale up and down your compute assets as needed.

Someone else manages the data center.

http://aws.amazon.com/education http://globus.org/ http://wiki.galaxyproject.org/Cloud







The Open Source Toolkit for Cloud Computing



### Instant CloudMan http://usegalaxy.org/cloudlaunch



### Galaxy is available: Commercial Support

A ready-to-use appliance (BioTeam)

Cloud-based solutions (ABgenomica, AIS, Appistry, GenomeCloud)

**Consulting & Customization** (Arctix, BioTeam, Deena Bioinformatics)











### **Galaxy Project: Further reading & Resources**

http://galaxyproject.org http://usegalaxy.org http://getgalaxy.org http://wiki.galaxyproject.org/Cloud http://bit.ly/gxychoices

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# **Basic Analysis**

# Which genes have most overlapping Repeats?

http://cloud1.galaxyproject.org/ http://cloud2.galaxyproject.org/ http://cloud3.galaxyproject.org/ http://cloud4.galaxyproject.org/

(~ http://usegalaxy.org/galaxy101)

# Genes & Repeats: A General Plan

- Get some data
  - Get Data → UCSC Table Browser
- Identify which genes/exons have Repeats
- Count Repeats per exon
- Visualize, save, download, ... exons with most Repeats

http://cloud1.galaxyproject.org/ http://cloud2.galaxyproject.org/ http://cloud3.galaxyproject.org/ http://cloud4.galaxyproject.org/

(~ http://usegalaxy.org/galaxy101 )



#### **Exons**



#### Repeats

(Identify which genes/exons have Repeats)







## Operate on Genomic Intervals → Join (Identify which genes/exons have Repeats)





#### **Exons**



#### **Overlap pairings**





Join, Subtract, and Group → Group (Count Repeats per exon)





#### Exon overlap counts

Exons

We've answered our question, but we can do better. Incorporate the overlap count with rest of Exon information





**Exons** 



#### Join on exon name

### Join, Subtract, and Group $\rightarrow$ Join

(Incorporate the overlap count with rest of Exon information)







### Text Manipulation $\rightarrow$ Cut

(Incorporate the overlap count with rest of Exon information)

# Genes & Repeats: Exercise

Include genes/exons with no overlaps in final output. Set the score for these to 0.

Everything you need will be in the toolboxes we used in the first Gene/Exon-Repeats exercise.

http://cloud1.galaxyproject.org/ http://cloud2.galaxyproject.org/ http://cloud3.galaxyproject.org/ http://cloud4.galaxyproject.org/

### **One Possible Solution**



Solution from Stanford Kwenda and Caron Griffiths in Pretoria. Takes advantage of the fact that Exons already have 0 scores.

### **Basic Analysis: Further reading & Resources**

http://usegalaxy.org/galaxy101 https://vimeo.com/76343659

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# NGS Data Quality Control

- FASTQ format
- Examine quality in an RNA-Seq dataset
- Trim/filter as we see fit, hopefully without breaking anything.

# Quality Control is not sexy. It is vital.

### What is **FASTQ**?

### Specifies sequence (FASTA) and quality scores (PHRED)

### • Text format, 4 lines per entry



### • FASTQ is such a cool standard, there are 3 (or 5) of them!

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33	59	)	64	73			104	126
S - Sanger	Phred+33,	93	values	(0,	93)	(0 to 60 expected	in raw reads)	
I - Illumina 1.3 X - Solexa	Phred+64, Solexa+64,	62 67	values values	(0, (-5,	62) 62)	(0 to 40 expected (-5 to 40 expected	in raw reads) d in raw reads)	

### http://en.wikipedia.org/wiki/FASTQ\_format

### **NGS Data Quality Exercise**

Create new history (cog) → Create New Get some data

Shared Data → Data Libraries

- → RNA-Seq Example\*
  - → Untrimmed FASTQ

→ Select MeOH\_REP1\_R1, MeOH\_REP1\_R2 and then Import to current history

**UCDAVIS** Bioinformatics Core

\* RNA-Seq example datasets from the 2013 UC Davis Bioinformatics Short Course. http://bit.ly/ucdbsc2013

### NGS Data Quality: Assessment tools

NGS QC and Manipulation → FastQC

Gives you a lot a lot of information but little control over how it is calculated or presented.

http://bit.ly/FastQCBoxPlot

### NGS Data Quality: Sequence bias at front of reads?



From a sequence specific bias that is caused by use of random hexamers in library preparation.

Hansen, et al., "Biases in Illumina transcriptome sequencing caused by random hexamer priming" *Nucleic Acids Research*, Volume 38, Issue 12 (2010)

### NGS Data Quality: Trim as we see fit

- Trim as we see fit: Option 1
  - NGS QC and Manipulation →
    FASTQ Trimmer by column
  - Trim same number of columns from every record
  - Can specify different trim for 5' and 3' ends



### NGS Data Quality: Base Quality Trimming

- Trim Filter as we see fit: Option 2
  - NGS QC and Manipulation →
    Filter FASTQ reads by quality
    score and length
  - Keep or discard whole reads
  - Can have different thresholds for different regions of the reads.
  - Keeps original read length.



### NGS Data Quality: Base Quality Trimming

- Trim as we see fit: Option 3
  - NGS QC and Manipulation →
    FASTQ Quality Trimmer by sliding window
  - Trim from both ends, using sliding windows, until you hit a high-quality section.
  - Produces variable length reads





### Trim? As we see fit?

- Introduced 3 options
  - One preserves original read length, two don't
  - One preserves number of reads, two don't
  - Two keep/make every read the same length, one does not
  - One preserves pairings, two don't
### Trim? As we see fit?

### Choice depends on downstream tools

- Find out assumptions & requirements for downstream tools and make appropriate choice(s) now.
- How to do that?
  - Read the tool documentation
  - http://biostars.org/
  - http://seqanswers.com/
  - http://galaxyproject.org/search





## NGS Data Quality: Base Quality Trimming



- I really want to use Option 3:
- NGS QC and Manipulation →
   FASTQ Quality Trimmer by sliding window

but ...

"Mixing paired- and single- end reads together is not supported." Tophat Manual

"If you are performing RNA-seq analysis, there is no need to filter the data to ensure exact pairs before running Tophat." Jen Jackson

Galaxy User Support Person Extraordinaire

"Dang."

Most of us

Running Tophat on *no-longer-cleanly-paired* data *does map the reads*, but, it no longer keeps track of read pairs in the SAM/BAM file.

## Keeping paired ends paired: Options

- Don't bother.
- Run a workflow that removes any unpaired reads before mapping.
- Run the Picard Paired Read Mate Fixer after mapping reads.
- Use sliding windows for QC, but keep empty reads.

## NGS Data Quality: Base Quality Trimming



I'll use Option 3 (*but* ...):

- NGS QC and Manipulation → FASTQ
   Quality Trimmer by sliding window
  - Check "Keep reads with zero length"

Run again:

 NGS QC and Manipulation → FastQC on trimmed dataset

### NGS Data Quality: Base Quality Trimming

Distribution of sequence lengths over all sequences Sequence Length 41 43 45 47 49 51 53 55 Sequence Length (bp)

New Problem? Now some reads are so short they are just noise and can't be meaningfully mapped

Option 2 can fix this (but break pairings).

Or, your mapper may have an option to ignore shorter reads

### NGS Data Quality: Sequencing Artifacts

Repeat this process with MeOH Rep1 R2 (the reverse reads) ... and there's a problem in Overrepresented sequences:

### Overrepresented sequences

Sequence	Count	Percentage	Possible Source
CTGTGTATTTGTCAATTTTCTTCTCCACGTTCTTCTCGGCCTGTTTCCGTAGCCT	590	0 3541692929220167	No Hit
TTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTT	342	0.2052981325073385	No Hit
CGGCCACAAATAAACACAGAAATAGTCCAGAATGTCACAGGTCCAGGGCAGAGGA	325	0.19509325457568719	No Hit
CTGCATTATAAAAAGGACAGCCAGATATCAACTGTTACAGAAATGAAAATAAGACG	230	0.13806599554587093	No Hit
CGGCCGCAAATAAACACAGAAATAGTCCAGAATGTCACAGGTCCAGGGCAGAGGA	199	0.11945710049403614	No Hit
GTCAGCTCAACTTGTAGGCCCCCAAAAGAAAACAGCGTCTTACTGGGGAGGGA	197	0.11825652661972422	No Hit

NGS QC and Manipulation  $\rightarrow$  Remove sequencing artifacts But this will break pairings.

### NGS Data Quality: Done with 1st Replicate!

Now, only 5 more to go!

Workflows?

Create a QC workflow that does the trimming

Or, cheat and import the Sliding window QC, paired end, keep empties published workflow

Or, really cheat and just import the already trimmed datasets from the RNA-Seq Example → Trimmed FASTQ shared data library

### NGS Data Quality: Further reading & Resources

### **FastQC Documenation**

Read Quality Assessment & Improvement by Joe Fass From the UC Davis 2013 Bioinformatics Short Course Manipulation of FASTQ data with Galaxy

by Blankenberg, et al.

# RNA-seq Exercise: Mapping with Tophat Cheat Alert!

We are going to talk about Tophat but we aren't going to run it today: 1. It takes a lot of time to run 2. Tophat2 has issues on these instances Therefore we will talk about Tophat, and then use results of Tophat run that was run before the workshop

**RNA-seq Exercise: Mapping with Tophat** 

Create a new history Import all datasets from library: RNA-Seq Example → Mapped Reads and genes\_chr12.gtf

### **RNA-seq Exercise: Mapping with Tophat**

- Tophat looks for best place(s) to map reads, and best places to insert introns
- Imagine pages and pages of discussion on the intricacies and pitfalls of RNA-seq mapping here
- Things like mean inner distance, how much guidance to give it when predicting junctions, indel and "coverage" search, tie-breaking, ...

(See 5 pages on this at the end of these slides)

**RNA-Seq Mapping With Tophat: Resources** 

<u>RNA-Seq Concepts, Terminology, and Work Flows</u> by Monica Britton

<u>Aligning PE RNA-Seq Reads to a Genome</u> by Monica Britton

both from the <u>UC Davis 2013 Bioinformatics Short Course</u>

<u>RNA-Seq Analysis with Galaxy</u> by <u>Jeroen F.J. Laros</u>, <u>Wibowo Arindrarto</u>, <u>Leon Mei</u>

from the GCC2013 Training Day

### **RNA-Seq Analysis with Galaxy**

by Curtis Hendrickson, David Crossman, Jeremy Goecks

from the <u>GCC2012 Training Day</u>

# Differential Gene Expresison: Cuffdiff?

- Part of the Tuxedo RNA-Seq Suite (as are Tophat and Bowtie)
- Widely used and widely installed on Galaxy instances

### NGS: RNA Analysis → Cuffdiff

# Cuffdiff

Cuffdiff uses FPKM/RPKM as a central statistic. Total # mapped reads heavily influences FPKM/RPKM. Can lead to challenges when you have very highly expressed genes in the mix.

# **Cuffdiff Alternatives**

Rapaport, *et al.*, "Comprehensive evaluation of differential gene expression analysis methods for RNA-seq data." *Genome Biology* 2013, 14:R95 doi:10.1186/gb-2013-14-9-r95

Reviews 7 packages

Each tool has it's own strengths and weaknesses. What's a biologist to do?

## Alternatives: What's a biologist to do?

Learn the strengths and weaknesses of the tools you have ready access to. Are they a good match for the questions you are asking?

If not, then research alternatives, identify good options and then work with your bioinformatics/systems people to get access to those tools.

DESeq is an R based differential expression analysis package where expression analysis is much more effectively isolated between features.

Takes a simple, tab delimited list of features and read counts across different samples. First, have to create that list.

### htseq-count

Is a tool that walks BAM files producing these lists

NGS: SAM Tools → htseq-count once for each BAM file

Join the HTSeq datasets together on gene name Cut out the duplicate gene name columns OR, just use the 6x DESeq Prep workflow

NGS: RNA Analysis → DE Seq

DESeq output is a list of genes, sorted by adjusted P value, with lowest P values listed first

How many genes have an adjusted P value < 0.05 ?

**Differential Expression: Reading & Resources** 

<u>Comprehensive evaluation of differential gene</u> <u>expression analysis methods for RNA-seq data</u> by Rapaport, *et al*.

**DESeq Reference Manual** 

DESeq Galaxy Wrapper by Nikhil Joshi

<u>htseq-count Galaxy Wrapper</u> by Lance Parsons

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**Galaxy Community Resources: Galaxy Biostar** Tens of thousands of users leads to a lot of questions. Absolutely have to encourage community support. Project traditionally uses mailing list Just moved the user support list to Galaxy Biostar, an online forum, that uses the Biostar platform



## https://biostar.usegalaxy.org/

Galaxy Community Resources: Mailing Lists http://wiki.galaxyproject.org/MailingLists

## Galaxy-Dev

Questions about developing for and deploying Galaxy High volume (5200 posts in 2013, 900+ members)

### Galaxy-Announce

Project announcements, low volume, moderated Low volume ( 47 posts in 2013, 3400+ members)

### Galaxy-User (deprecated)

Questions about using Galaxy and usegalaxy.org High volume (1328 posts in 2013, 2600+ members)

## Unified Search: http://galaxyproject.org/search

# Coogle" Custom Search Search the entire set of Galaxy web sites and mailing lists using Google. Run this search at Google.com (useful for bookmarking) Want a different search? Project home



## Community: Public Galaxy Instances http://bit.ly/gxyServers

### **Interested in:**

ChIP-chip and ChIP-seq? ✓ Cistrome, Nebula **Statistical Analysis?** ✓ Genomic Hyperbrowser Protein synthesis? ✓ GWIPS-viz de novo assembly? ✓ CBIIT Galaxy **Reasoning with ontologies?** ✓ GO Galaxy **Repeats!** ✓ RepeatExplorer

Over 50 public Galaxy servers

## http://wiki.galaxyproject.org



### Contribute

Develop • Share **Issues & Requests** Teach • Support

Galaxy Project

Home • About

### Community & Project

Galaxy has a large and active user community and many ways to get involved.

Community

 Users: Share your histories, workflows, visualizations, data libraries, and Galaxy Pages, enabling others to use and learn from them

Contribute

### **Events**

### News

### Galaxy Event Horizon

Events with Galaxy-related content are listed here.

Also see the Galaxy Events Google Calendar for a listing of events and deadlin Community. This is also available as an RSS feed .

If you know of any event that should be added to this page and/or to the Galaxy Even send it to soutreach@glaxyproject.org .

For events prior to this year, see the Events Archive.

### Upcoming Events



Date	Topic/Event	Ver
May 6-7	Scaling Galaxy for Big Data	
May 9	Introduction to Galaxy Workshop	The (TG
May 12	Galaxy Workshop	Uni UK
	Galaxy Project Update	5th Mee Edir
May 13	Galaxy Workshop	Inst Med
May 12-14	Short course on RNA-seq and ChIP-seq	Uni Nor
May 16	Galaxy Initiation	For Plat Biol
May 19	Initiation au traitement et à l'analyse des données métabolomiques sur la plateforme scientifique web Galaxy IFB-MetaboHUB	8e I Lyo

### News Items

May 2014 Galaxy News



The May 2014 Galaxy Update Newsletter is out! There's a lot going on in the project and the community right now. The big news in the past month is the move from the Galaxy-User mailing list to Galaxy Biostar for user support. This has been running for a week now, and has been very well received.

The other big news is upcoming events. **Early registration for GCC2013 closes May 23**. Register now and save more than 70% on registration costs, and Training Day registration is an additional 55% off if you register for both at the same time. We are also pleased to announce this year's keynote speaker and the first ever GCC Hackathon.

There's also a Galaxy UK Tour which is visiting Norwich and Edinburgh in May, and there are at least 17 other Galaxy related events in the next 70 days in Norway, France, *online*, Croatia, Thailand, Canada, the US, the Netherlands, and Australia

As always, there are new papers (47 of them, including four we highlighted), new public Galaxy Servers (Globus Genomics Proteomics and SunLab Galaxy), new jobs (7 postings in 6 countries), new tools in the project ToolShed (um, *lots*), and a new public ToolShed (at the Dutch Techcentre for Life Sciences (DTL).

Dave Clements and the Galaxy Team

Posted to the Galaxy News on 2014-04-30

### Galaxy Biostar Launched

### Galaxy has teamed up with Biostar to create a Galaxy User support forum at https://biostar.usegalaxy.org!

We want to create a space where researchers using Galaxy can come together and share both scientific advice and practical tool help. Whether on usegalaxy.org, a CloudMan instance, or any other Galaxy (public or local), if you have something to say about *Using Galaxy*, this is the place to do it!

### Current integration with usegalaxy.org

- We imported the whole history of the galaxy-user@bx.psu.edu mailing list into Galaxy Biostar. Your prior posts are automatically claimed when you login!
- If you access Galaxy Biostar from http://usegalaxy.org (Menu: Help → Galaxy Biostar) you will be automatically logged in. A Galaxy Biostar account will be created for you if it did not previously exist. To obtain this account's password please use the password reset feature of Galaxy Biostar.
- When you have a question, search Galaxy Biostar directly from any Galaxy tool page.

Read more about how to get started on the Biostar wiki page.











# BALTIMORE, MD | JUNE 30 - JULY 2, 2014

# http://bit.ly/gcc2014







### **Galaxy Resources & Community: Videos**



"How to" screencasts on using and deploying Galaxy

Talks from previous meetings.

### http://vimeo.com/galaxyproject

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- 12:40 Project and Community Overview
- 13:00 Done (almost)

## The Galaxy Team



Enis Afgan

Dannon Baker

Dan Blankenberg

**Dave Bouvier** 

Marten Cech

John Chilton



**Dave Clements** 

Nate Coraor

**Carl Eberhard** 

**Dorine Francheteau** 

Jeremy Goecks

Sam Guerler



Jen Jackson



Greg von Kuster

Ross Lazarus

Anton Nekrutenko



**Nick Stoler** 

**James Taylor** 

http://wiki.galaxyproject.org/GalaxyTeam

### Galaxy is hiring post-docs and software engineers



Please help. http://wiki.galaxyproject.org/GalaxylsHiring

### Also Thanks To





B





Wellcome Trust Centre for Cell Biology



**Edinburgh Bioinformatics** 

Al Kerr Mick Watson James Pendergast





The Genome Analysis Centre<sup>™</sup>

# Feedback

# We need it!

http://bit.ly/gxyedifeedback
# Thanks

Dave Clements Galaxy Project Johns Hopkins University outreach@galaxyproject.org

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# Sharing, Publishing, and Reproducibility

# More Galaxy Terminology

# Share:

Make something available to someone else Publish:

Make something available to everyone

# Galaxy Page:

Analysis documentation within Galaxy; easy to embed any Galaxy object

Let's all share...

# Sharing & Publishing enables Reproducibility

Reproducibility: Everybody talks about it, but ...

Galaxy aims to push the goal of reproducibility from the bench to the bioinformatics realm

All analysis in Galaxy is recorded without any extra effort from the user.

Histories, workflows, visualizations and *pages* can be shared with others or published to the world.

# Sharing & Publishing enables Reproducibility



# Windshield splatter analysis with the Galaxy metagenomic pipeline

Sergei Kosakovsky Pond<sup>1,2,6,9</sup>, Samir Wadhawan<sup>3,6,7</sup>, Francesca Chiaromonte<sup>4</sup>, Guruprasad Ananda<sup>1,3</sup>, Wen-Yu Chung<sup>1,3,8</sup>, James Taylor<sup>1,5,9</sup>, Anton Nekrutenko<sup>1,3,9</sup> and The Galaxy Team<sup>1</sup>

#### **OPEN ACCESS ARTICLE**

illumina<sup>\*</sup>

#### **This Article**

Published in Advance October 9, 2009, doi: 10.1101/gr.094508.109

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#### - » Abstract Free

#### Current Issue

Search for Keyword: Advanced Search

October 2010, 20 (10)

Go



Apply today for the

Cancer GWAS Grant.

#### Footnotes

[Supplemental material is available online at http://www.genome.org. All data and tools described in this manuscript can be downloaded or used directly at http://galaxyproject.org. Exact analyses and workflows used in this paper are available at http://usegalaxy.org/u/aun1/p/windshield-splatter.] 🗧 Galaxy

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Using

Published Pages | aun1 | Windshield Splatter

# Windshield splatter analysis with the Galaxy metagenomic pipeline: A live supplement

SERGEI KOSAKOVSKY POND<sup>1,2,\*</sup>, SAMIR WADHAWAN<sup>3,6\*</sup>, FRANCESCA CHIAROMONTE<sup>4</sup>, GURUPRASAD ANANDA<sup>1,3</sup>, WEN-YU CHUNG<sup>1,3,7</sup>, JAMES TAYLOR<sup>1,5</sup>, ANTON NEKRUTENKO<sup>1,3</sup> and THE GALAXY TEAM<sup>1\*</sup>

Correspondence should addressed to SKP, JT, or AN.

#### How to use this document

This document is a live copy of supplementary materials for <u>the manuscript</u>. It provides access to the **exact** analyses and workflows discussed in the paper, so you can play with them by re-running, changing parameters, or even applying them to your own data. Specifically, we provide the two histories and one workflow found below. You can view these items by clicking on their name to expand them. You can also import these items into your Galaxy workspace and start using them; click on the green plus to import an item. To import workflows you must <u>create a Galaxy account</u> (unless you already have one) – a hassle-free procedure where you are only asked for a username and password.

This is the Galaxy history detailing the comparison of our pipeline to MEGAN:



This is the Galaxy history showing a generic analysis of metagenomic data. (This corresponds to the "A complete metagenomic pipeline" section of the manuscript and Figure 3A):



Galaxy History | metagenomic analysis



Galaxy Workflow | metagenomic analysis
 Generic workflow for performing a metagenomic analysis on NGS data.
 Generic workflow for performing a metagenomic analysis on NGS data.
 Generic workflow for performing a metagenomic analysis on NGS data.
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 Generic workflow for performing a metagenomic analysis on NGS data.

#### Accessing the Data

Windshield Splatter datasets analyzed in this manuscript can be accessed through this Galaxy Library. From

### http://usegalaxy.org/u/aun1/p/windshield-splatter





aun1

#### **Related Pages**

About this Page

All published pages Published pages by aun1

#### Rating

Community (6 ratings, 5.0 average)





>

### **Sharing for Galaxy Administrators Too**

Data Libraries Make data easy to find

Genome Builds

Care about a particular subset of life?

Galaxy Tool Shed

Wrapping tools and datatypes

RNA-seq Exercise: Mapping with Tophat

Yes, but how *might* we run Tophat?

 Tophat looks for best place(s) to map reads, and best places to insert introns

 Imagine pages and pages of discussion on the intricacies and pitfalls of RNA-seq mapping here. Mapping with Tophat: mean inner distance

Expected distance between paired end reads

- Determined by sample prep
- We'll use 90\* for mean inner distance
- We'll use 50 for standard deviation

\* The library was constructed with the typical Illumina TruSeq protocol, which is supposed to have an average insert size of 200 bases. Our reads are 55 bases (R1) plus 55 bases (R2). So, the Inner Distance is estimated to be 200 - 55 - 55 = 90

From the 2013 UC Davis Bioinformatics Short Course

Mapping with Tophat: Use Existing Annotations?

You can bias Tophat towards known annotations

- Use Own Junctions → Yes
  - Use Gene Annotation → Yes
  - Gene Model Annotation → genes\_chr12.gtf
- Use Raw Junctions → Yes (tab delimited file)
- Only look for supplied junctions → Yes

### Mapping with Tophat: Make it quicker?

#### Warning: Here be dragons!

#### Allow indel search → No

#### ● Use Coverage Search → No (wee dragons)

TopHat generates its database of possible splice junctions from two sources of evidence. The first and strongest source of evidence for a splice junction is when two segments from the same read (for reads of at least 45bp) are mapped at a certain distance on the same genomic sequence or when an internal segment fails to map - again suggesting that such reads are spanning multiple exons. With this approach, "GT-AG", "GC-AG" and "AT-AC" introns will be found *ab initio*. The second source is pairings of "coverage islands", which are distinct regions of piled up reads in the initial mapping. Neighboring islands are often spliced together in the transcriptome, so TopHat looks for ways to join these with an intron. We only suggest users use this second option (--coverage-search) for short reads (< 45bp) and with a small number of reads (<= 10 million). This latter option will only report alignments across "GT-AG" introns

TopHat Manual

Mapping with Tophat: Max # of Alignments Allowed Some reads align to more than one place equally well. For such reads, how many should Tophat include? If more than the specified number, Tophat will pick those with the best mapping score.

Tophat break ties randomly.

#### Tophat assigns equal fractional credit to all n

Instructs TopHat to allow up to this many alignments to the reference for a given read, and choose the alignments based on their alignment scores if there are more than this number. The default is 20 for read mapping. Unless you use --report-secondary-alignments, TopHat will report the alignments with the best alignment score. If there are more alignments with the same score than this number, TopHat will randomly report only this many alignments. In case of using --report-secondaryalignments, TopHat will try to report alignments up to this option value, and TopHat may randomly output some of the alignments with the same score to meet this number.

**TopHat Manual** 

Mapping with Tophat: Lets do it some more! NGS: RNA Analysis  $\rightarrow$  Tophat for the remaining replicates Or not.

# Some Galaxy Terminology

### Dataset:

Any input, output or intermediate set of data + metadata History:

A series of inputs, analysis steps, intermediate datasets, and outputs

### Workflow:

A series of analysis steps Can be repeated with different data

### Exons and Repeats *History* → Reusable *Workflow*?

- The analysis we just finished was about
  - Human chr22
  - Overlap between exons and Repeats
- But, ...
  - there is nothing inherent in the analysis about humans, exons or repeats
  - It is a series of steps that sets the score of one set of features to the number of overlaps from another set of features.

## Create a Workflow from a History

#### **Extract Workflow from history**

Create a workflow from this history. Edit it to make some things clearer.

 $(cog) \rightarrow Extract Workflow$ 

Run / test it Guided: rerun with same inputs Did that work?

#### On your own:

Count # of exons in each Repeat Did that work? *Why not?* Edit workflow: doc assumptions

Histor	v 2 🌣
impc 33.3	HISTORY LISTS Saved Histories Histories Shared with Me
22: C	CURRENT HISTORY
data	Create New
FPKN	Copy History
21: C data diffe	Copy Datasets Share or Publish
<u>20: C</u>	Dataset Security
data	Resume Paused Jobs
track	Collapse Expanded Datasets
<u>19: C</u>	Include Deleted Datasets
data	Include Hidden Datasets
diffe	Unhide Hidden Datasets
<u>18: C</u>	Purge Deleted Datasets
data	Show Structure
FPKN	Export to File
<u>17: C</u>	Delete
data	Delete Permanently
diffe	OTHER ACTIONS
<u>16: C</u> data trackin	Import from File

### Community can create, vote and comment on issues



### http://bit.ly/gxytrello

# Galaxy Australasia • • 20 1 Workshop • 4

We also support community organized efforts and events.



### Galaxy Resources & Community: CiteULike Group

Over

1500

papers

17 tags

#### citeulike 🗉

CiteULike MyCiteULike Group: Galaxy	₽ <u>Search</u>	Logged in as galaxyproject	Log Out
Group: Galaxy - library 1437 articles         You are an administrative member of this group.         Invite other CiteULike users to join, or invite people who don't use CiteULike yet.	collaborativ	e cyberin methods workbench usemain tools isgalaxy cloud shared unknown uselocal project howto reproducibility other usepublic refpublic visualization usecloud	697 466 108 91 80 50 50 47 37 32 30 28 23 19 12 7 3
<ul> <li>Detection of PIWI and pIRNAs in the mitochondria of mammalian cancer cells Biochemical and Biophysical Research Communications (March 2014), doi:10.1016/j.bbrc.2014.02.112 by ChangHyuk Kwon, Hyosun Tak, Mina Rho, et al. posted to methods by galaxyproject to the group Galaxy keyed Kwon2014Detection on 2014-03-04 18:53:07 **/ al Copy My Copy</li> <li>CanSNPer: a hierarchical genotype classifier of clonal pathogens Bioinformatics (25 February 2014), doi:10.1093/bioinformatics/btu113 by Adrian Lärkeryd, Kerstin Myrtennäs, Edvin Karlsson, et al. posted to to tools by galaxyproject to the group Galaxy keyed Larkeryd2014CanSNPer on 2014-03-04 18:51:21 **/ Abstract Copy My Copy</li> <li>Web-based Workflow Planning Platform Supporting the Design and Execution of Complex Multiser</li> </ul>	long with 1 per	rson	

pp. 1-1, doi:10.1109/jbhi.2013.2297167

