

Introduction to Galaxy

Saint Louis University

St. Louis, Missouri

April 30, 2013

Dave Clements, Emory University

<http://galaxyproject.org/>



SAINT LOUIS UNIVERSITY



 Galaxy



Edward A. Doisy Department of
Biochemistry and Molecular Biology

at Saint Louis University School of Medicine



SAINT LOUIS
UNIVERSITY

Agenda

- 9:00 **Welcome**
- 9:20 Basic Analysis with Galaxy
- 10:30 Basic Analysis into Reusable Workflows
- 11:00 RNA-Seq Example Part I
- 11:30 Lunch (on your own)
- 1:00 RNA-Seq Example Part II
- 1:30 Galaxy Project Overview
- 2:00 RNA-Seq Example Part III
- 2:30 Sharing, Publishing and Reproducibility
- 2:50 Break
- 3:10 ChIP-Seq Example
- 5:00 Done

Introductions

In 60 seconds or less tell us

- your name
- your affiliation(s)
- something about your research
- something about what you want to learn

Goals

1. Introduce Galaxy
2. Introduce bioinformatics concepts and formats
3. Hands-on experience
 - Load and integrate data
 - Perform bioinformatic analysis with Galaxy
 - Save, share describe and publish your analyses
 - Visualize your results
 - Set up your own Galaxy server in the cloud

This workshop will not cover details of how tools are implemented, or new algorithm designs, or which assembler or mapper or ... is best for you.

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Basic Analysis

On human chromosome 22,
which coding exons have the most
repeats in them?

<http://cloud1.galaxyproject.org/>

<http://cloud2.galaxyproject.org/>

<http://cloud3.galaxyproject.org/>

(~ <http://usegalaxy.org/galaxy101>)

Exons & Repeats: A General Plan

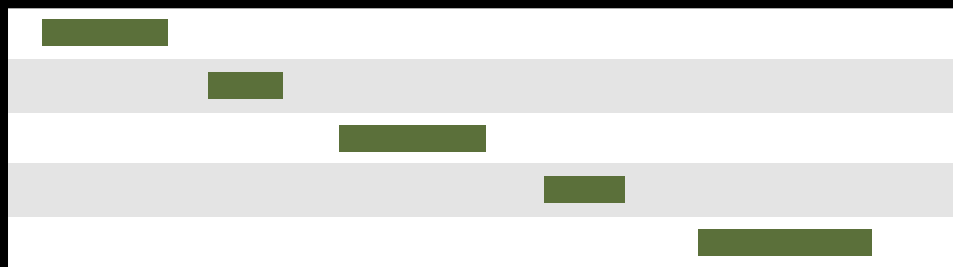
- Get some data
 - Coding exons on chromosome 22
 - Repeats on chromosome 22
- Mess with it
 - Identify which exons have repeats
 - Count repeats per exon
 - Save, download, ... exons with most repeats

<http://cloud1.galaxyproject.org/>

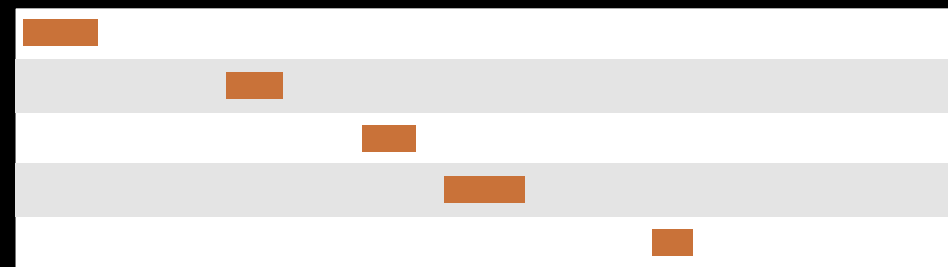
<http://cloud2.galaxyproject.org/>

<http://cloud3.galaxyproject.org/>

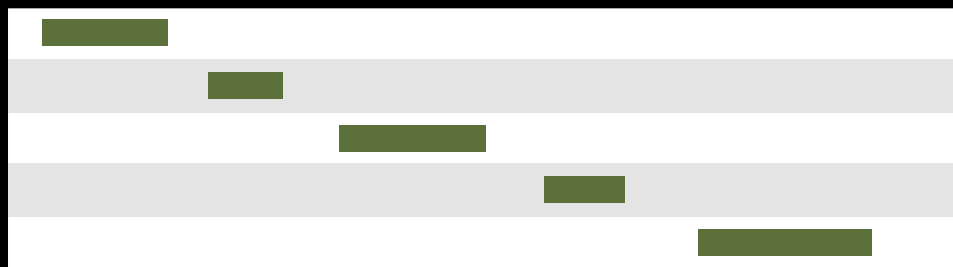
(~ <http://usegalaxy.org/galaxy101>)



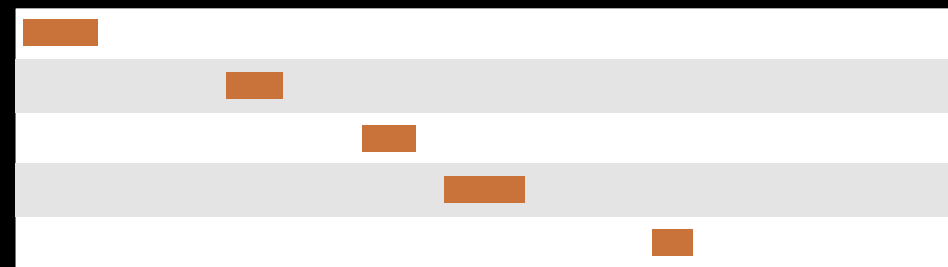
Exons, from UCSC



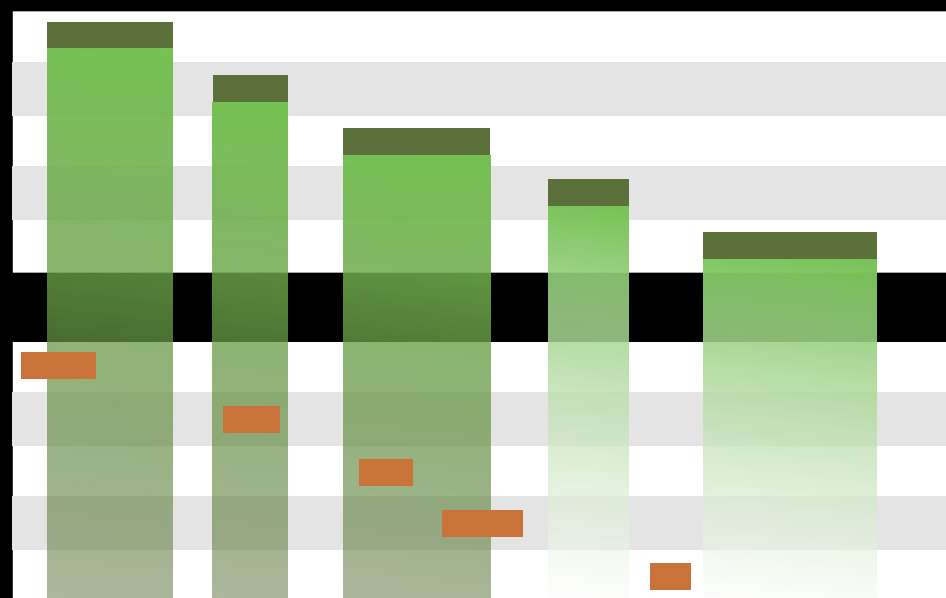
Repeats, from UCSC



Exons, from UCSC



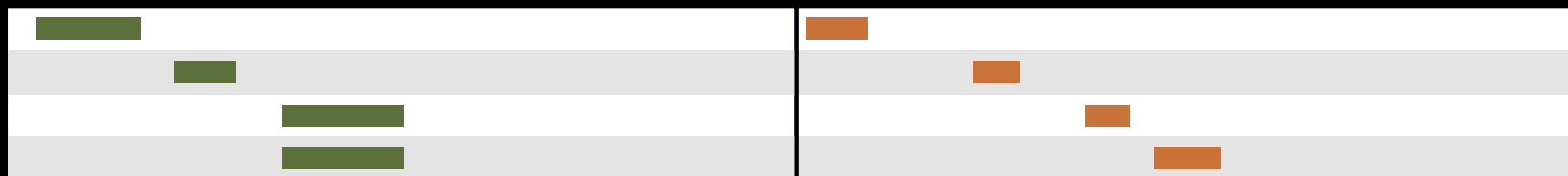
Repeats, from UCSC

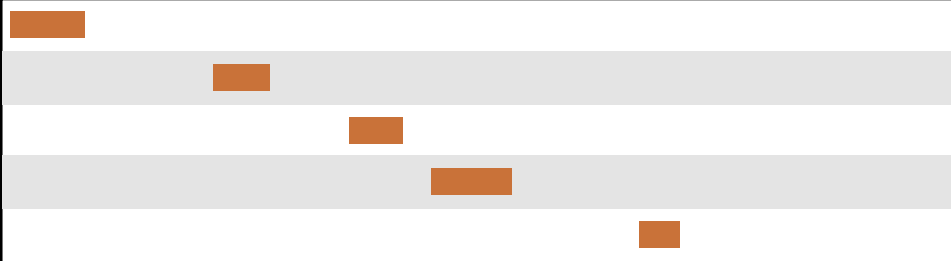
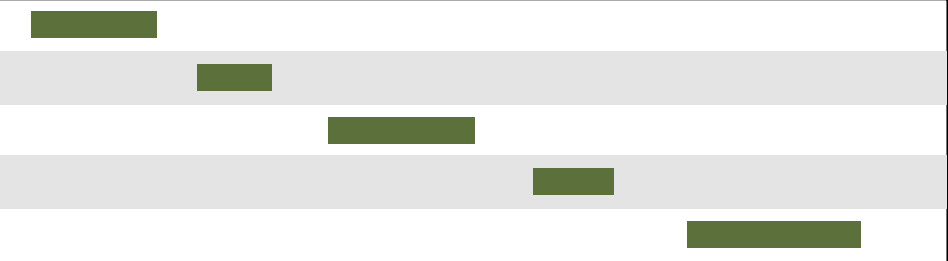


Exons, from UCSC

Repeats, from UCSC

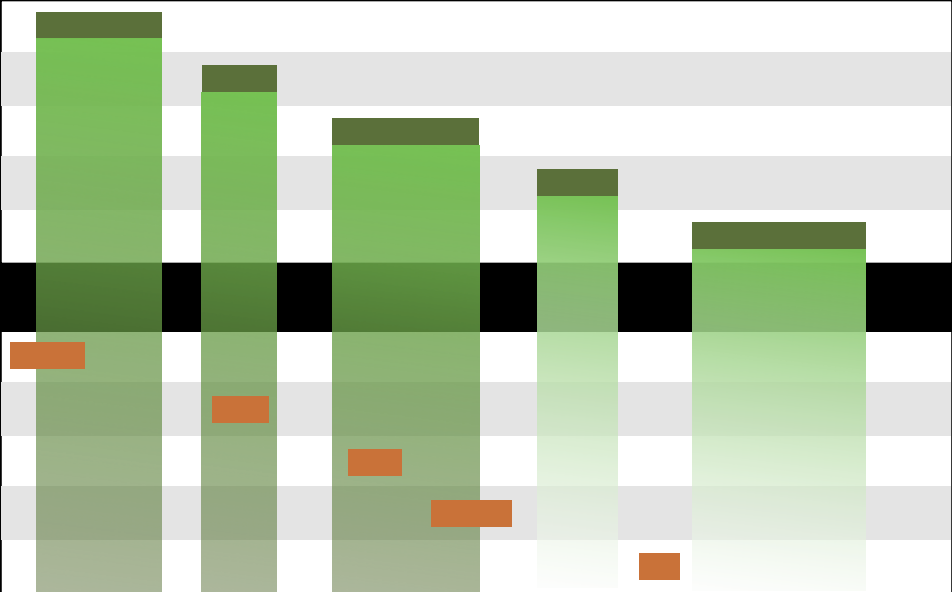
Overlap pairings





Exons, from UCSC

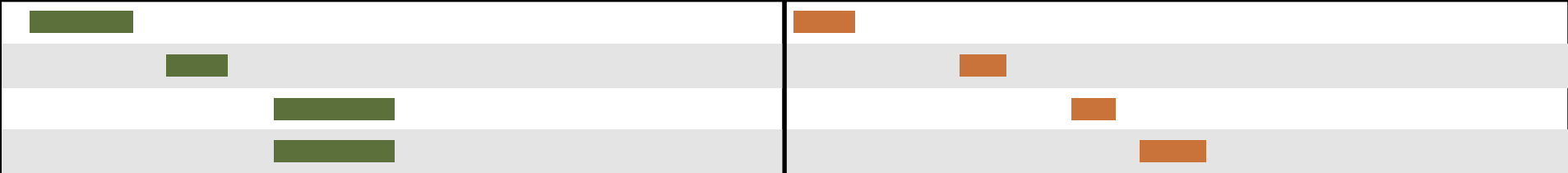
Repeats, from UCSC



Exons, from UCSC

Repeats, from UCSC

Overlap pairings

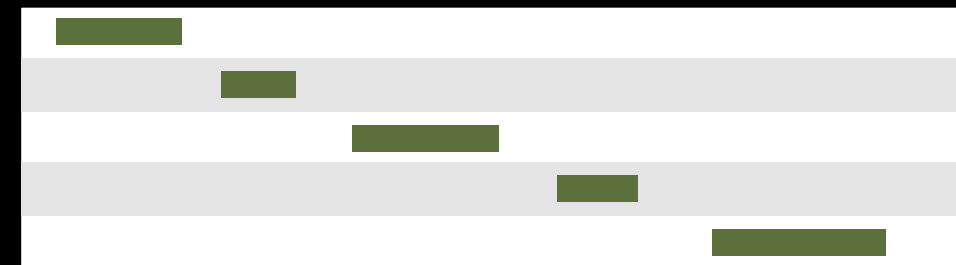


	1
	1
	2

Exon overlap counts



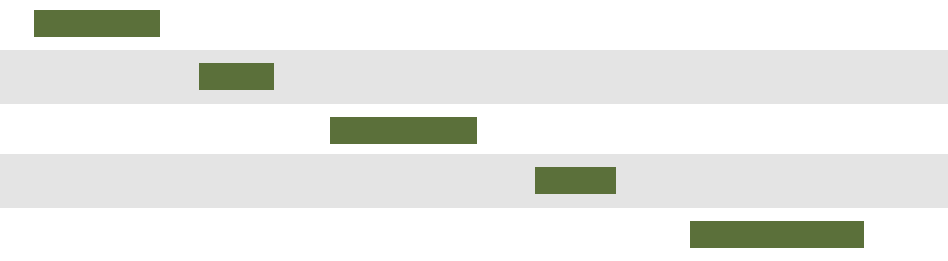
Exon overlap counts









Exons, from UCSC

	1
	1
	2

Exon overlap counts






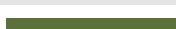
Exons, from UCSC

	1		0
	1		0
	2		0

Join on exon name

	1
	1
	2




Exon overlap counts

Exons, from UCSC

	1		0
	1		0
	2		0

Join on exon name

	1
	1
	2

Rearrange columns w/
cut

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Some Galaxy Terminology

Dataset:

Any input, output or intermediate set of data + metadata

History:

A series of inputs, analysis steps, intermediate datasets, and outputs

Workflow:

A series of analysis steps

Can be repeated with different data

Exons and Repeats *History* → Reusable *Workflow*?

- The analysis we just finished was about
 - Human chromosome 22
 - Overlap between exons and repeats
- But, ...
 - there is **nothing inherently** in the analysis **about humans, chromosomes, exons or repeats**
 - It is a series of steps that **sets the score of one set of features to the number of overlaps from another set of features.**

Create a generic *Overlap* Workflow

Extract Workflow from history

Create a workflow from this history.
Edit it to make some things clearer.

Run / test it

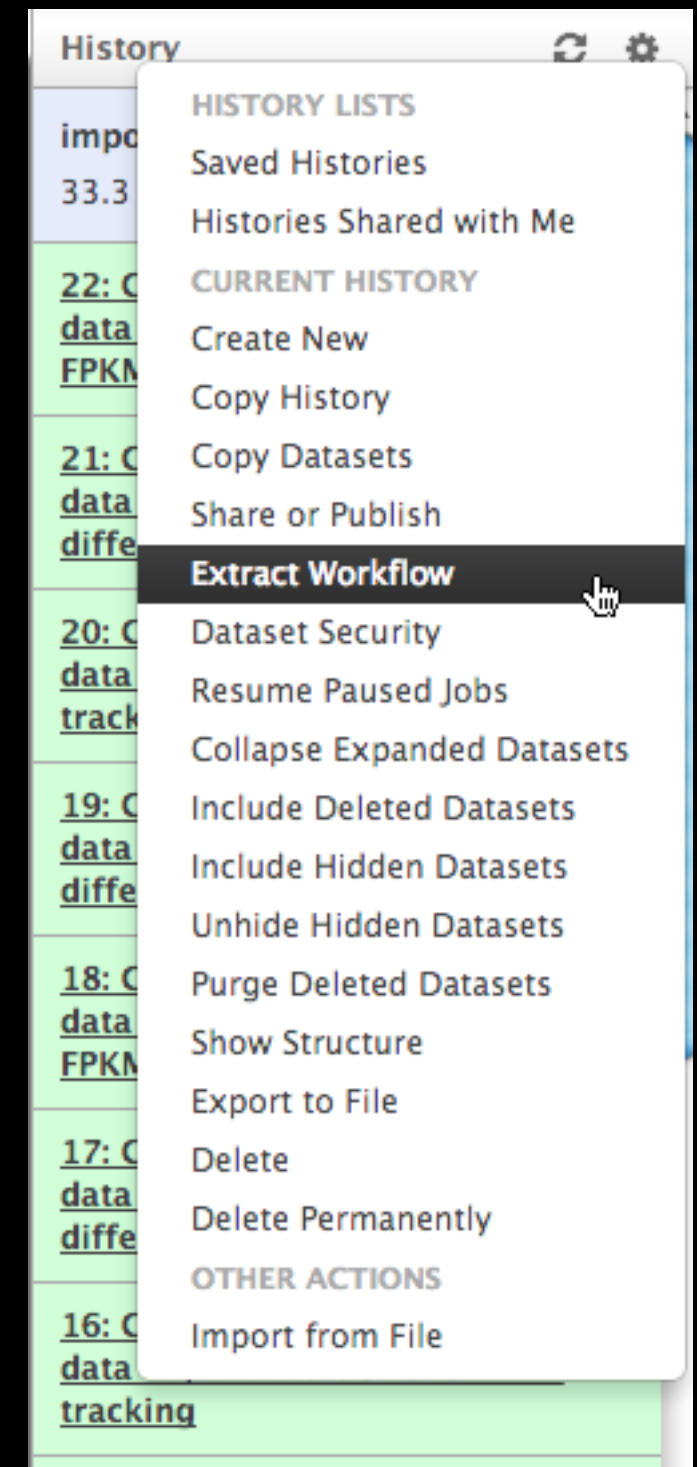
Guided: rerun with same inputs

On your own:

Count # CpG islands in each exon
Did that work?

On your own:

Count # of exons in each repeat
Did that work? *Why not?*
Edit workflow: doc assumptions



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RNA-seq Exercise

Shared Data → Published Pages

→ RNA-Seq Analysis Exercise

RNA-seq Exercise: A Plan

- Get input datasets; hg19, will mostly map to chr19
- Look at quality
- Trim as we see fit.
- Map the reads to the human reference using Tophat
- Run Cufflinks on Tophat output to assemble reads into transcripts
- Visualize it

RNA-seq Exercise: A Plan

- Get input datasets; hg19, will mostly map to chr19
 - All datasets are FASTQ and from the Body Map 2.0 project
 - Shared Data → Published Pages → RNA-Seq Analysis Exercise
 - or Shared Data → Data Libraries → RNA-Seq Example

What is FASTQ?

- Specifies sequence (FASTA) and quality scores (PHRED)
- Text format, 4 lines per entry

```
@SEQ_ID
GATTTGGGGTTCAAAGCAGTATCGATCAAATAGTAAATCCATTTGTTCAACTCACAGTTT
+
! ' ' * ( ( ( ( * * * + ) ) % % % + + ) ( % % % % ) . 1 * * * - + * ' ' ) ) * * 55CCF>>>>>>CCCCCCC65
```

- FASTQ is such a cool standard, there are 3 (or 5) of them!

[illegible]

http://en.wikipedia.org/wiki/FASTQ_format

RNA-seq Exercise: A Plan

Look at quality Options 1 & 2:

1. NGS QC and Manipulation → Compute Quality Statistics
and then Draw quality score boxplot

No control over how it is calculated or presented.

2. NGS QC and Manipulation → FastQ Summary Statistics,
Graph / Display Data → Boxplot of quality statistics

Lots of control over what the box plot looks like, but no
additional information; stats in text and graphic formats

RNA-seq Exercise: A Plan

- Look at quality: Option 3
 - NGS QC and Manipulation → **FastQC**
 - Gives you a lot a lot more information but little control over how it is calculated or presented.
- This is what we have done

RNA-seq Exercise: A Plan

RNA-seq Exercise: A Plan

*“For the love of all that is **holy**, please trim your reads!”*

Chris Mason, ABRF NGS Study Report, March 4, 2013

RNA-seq Exercise: A Plan

“For the love of all that is holy, please trim your reads!”

Chris Mason, ABRF NGS Study Report, March 4, 2013

- Trim as we see fit: Option 1
 - **NGS QC and Manipulation** → **FASTQ Trimmer by column**
 - Trim same number of columns from every record
 - Can specify different trim for 5' and 3' ends

RNA-seq Exercise: A Plan

- Look at quality
- ~~Trim~~ Filter as we see fit: Option 2
 - NGS QC and Manipulation → **Filter FASTQ reads by quality score and length**
 - Keep or discard whole reads at a time
 - Can have different thresholds for different regions of the reads.
 - Keeps original read length.

<http://bit.ly/GxyRNASeqEx>

RNA-seq Exercise: A Plan

- Look at quality
- Trim as we see fit: Option 3
 - NGS QC and Manipulation → **FASTQ Quality Trimmer by sliding window**
 - Trim from both ends, using sliding windows, until you hit a high-quality section.
 - **Produces variable length reads**

Trim? *As we see fit?*

- Introduced 3 options
 - One preserves original read length, two don't
 - One preserves number of reads, two don't
 - Two keep/make every read the same length, one does not
 - One preserves pairings, two don't
 - Options are not mutually exclusive!

Trim? *As we see fit?*

- Choice depends on downstream tools
- Find out assumptions & requirements for downstream tools and make appropriate choice(s) now.
- How to do that?
 - <http://biostars.org/>
 - <http://seqanswers.com/>
 - <http://galaxyproject.org/search>



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RNA-seq Exercise: A Plan

- Get input datasets; hg19, will mostly map to chr19
- Look at quality
- Trim as we see fit.
- Map the reads to the human reference using Tophat
 - *Imagine pages and pages of discussion on the intricacies and pitfalls of RNA-seq mapping here.*

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What is Galaxy?

- **A free (for everyone) web service** integrating a wealth of tools, compute resources, terabytes of reference data and permanent storage
- **Open source software** that makes integrating your own tools and data and customizing for your own site simple
- These options result in several **ways to use Galaxy**

<http://galaxyproject.org>

Galaxy is available ...

- As a free (for everyone) web service

<http://usegalaxy.org>

However, *a centralized solution cannot scale to meet the analysis needs of the entire world.*

Galaxy is available ...

- As a free (for everyone) web service

<http://usegalaxy.org>

- As open source software

<http://getgalaxy.org>

As Open Source Software: Local Galaxy Instances

- Galaxy is designed for local installation and customization
- Easily integrate new tools
- Easy to deploy and manage on nearly any (unix) system
- Run jobs on existing compute clusters
- Requires a computational resource on which to be deployed

<http://getgalaxy.org>

Encourage **Local** Galaxy Instances

- Encourage and support Local Galaxy Instances
- Support **increasingly decentralized model** and improve access to existing resources
- Focus on building **infrastructure to enable the community to integrate and share** tools, workflows, and best practices

Galaxy Tool Shed

<http://toolshed.g2.bx.psu.edu>

The screenshot shows the Galaxy Tool Shed interface for the 'clustalomega' repository. The left sidebar contains links for 'Repositories', 'Browse by category', 'Browse all repositories', and 'Login to create a repository'. The main content area displays the 'Repository revision' section with a dropdown menu showing '2:bb1847435ec1'. Below this, the 'clustalomega' repository details are shown, including a 'Clone this repository' link, the repository name, a synopsis, a detailed description, the revision number, the owner, and the number of times it has been downloaded. A table at the bottom lists the tools available in the repository.

name	description	version	requirements
Clustal Omega	multiple sequence alignment program for proteins	1.0.2	none

The screenshot shows the Galaxy Tool Shed interface with a list of repositories. The left sidebar is the same as in the previous screenshot. The main content area displays a table of repositories with columns for Name, Synopsis, Revision, Category, and Owner. The table lists several repositories, including 'abyss_toolsuite', 'agile_wrapper', 'asdf', 'assemblystats', and 'bam_to_bigwig'.

Name	Synopsis	Revision	Category	Owner
abyss_toolsuite	This suite contains Abyss and Abyss-PE config files and wrappers for Galaxy	0:92636934a189	Assembly	edward-kirton
agile_wrapper	Quickly match reads to a reference genome or sequence file	0:d6a426afaa46	Next Gen Mappers, Sequence Analysis	simonl
asdf	asdf	-1:000000000000	Statistics, Text Manipulation	vivek
assemblystats	Summarise an assembly (e.g. N50 metrics)	0:6544228ea290	Next Gen Mappers, Sequence Analysis	konradpaszkiewicz
bam_to_bigwig	Generate BigWig coverage files from BAM files. Allows gapped reads to be split (useful for RNA-Seq).	5:5b40b93ebae3	Convert Formats, SAM, Visualization	lparsons

Encourage **Public** Galaxy Instances

<http://bit.ly/gxyServers>

Interested in:

ChIP-chip and ChIP-seq?

✓ Cistrome

Statistical Analysis?

✓ Genomic Hyperbrowser

Protein synthesis?

✓ GWIPS-viz

de novo assembly?

✓ CBIIT Galaxy

Reasoning with ontologies?

✓ OPPL Galaxy

Repeats!

✓ RepeatExplorer

Everything?

✓ Andromeda

Plus many more

As Open Source Software: Local Galaxy Instances

- Galaxy is designed for local installation and customization
- Easily integrate new tools
- Easy to deploy and manage on nearly any (unix) system
- Run jobs on existing compute clusters
- Requires a **computational resource** on which to be deployed

<http://getgalaxy.org>

Got your own cluster?

- Galaxy **works with any DRMAA** compliant cluster job scheduler (which is most of them).
- Galaxy is **just another client** to your scheduler.



Galaxy is available ...

- As a free (for everyone) web service

<http://usegalaxy.org>

- As open source software

<http://getgalaxy.org>



- *On the Cloud*

<http://usegalaxy.org/cloud>

We are using this right now, and you will set up your own instance today

<http://aws.amazon.com/education>

Galaxy Resources and Community

Mailing Lists (very active)

Unified Search

Issues Board

Events Calendar, News Feed

Community Wiki

GalaxyAdmins

Screencasts

Tool Shed

Public Installs

CiteULike group, Mendeley mirror

Annual Community Meeting

<http://wiki.galaxyproject.org>

Galaxy Resources and Community: Mailing Lists

<http://wiki.galaxyproject.org/MailingLists>

Galaxy-Announce

Project announcements, low volume, moderated

Low volume (42 posts, 1600 members in 2012)

Galaxy-User

Questions about using Galaxy and usegalaxy.org


High volume (2900 posts, 2700 members in 2012)

Galaxy-Dev

Questions about developing for and deploying Galaxy

High volume (4500 posts, 850 members in 2012)

Unified Search: <http://galaxyproject.org/search>

 **Galaxy Web Search**

Search the entire set of Galaxy web sites and mailing lists using Google.

[Run this search at Google.com \(useful for bookmarking\)](#)

Want a [different search](#)?

[Project home](#)

Find

Everything on ...

Tools for ...

Email about ...


Source code for ...

Published Histories, Pages, Workflows, about ...

Documentation on ...

Papers using Galaxy for ...

Related feature requests

 **Galaxy Web Search**

About 444 results (0.06 seconds)

[Galaxy | Accessible Data | ChIP-seq exercise](#)

Community can create, vote and comment on issues

The screenshot displays a Trello board titled "Galaxy: Development Inbox" with a "Public" status. The board is organized into four main columns: "Inbox", "Developer ideas", "Bug Reports", and "Issues from Bitbucket".

- Inbox:** Contains five cards. The first card is a guide on adding cards. The second card discusses a filter and sort issue. The third card reports a problem with uploaded fastq file datatypes. The fourth card is a reference genome request. The fifth card is a feature request for manually hiding datasets.
- Developer ideas:** Contains five cards. The first is about anonymous use of workflows. The second is a feature request to restart failed workflows. The third is about Google Drive/Dropbox/Box integration. The fourth is a bug report about importing deleted datasets. The fifth is a request for a standalone web application for visualizations.
- Bug Reports:** Contains five cards. The first is about workflow step hiding. The second is about a broken workflow view. The third is about running jobs with limits. The fourth is about a FASTQ summary statistics tool tip. The fifth is about a bug when using data_column.
- Issues from Bitbucket:** Contains five cards. The first is about disabling automatic history creation. The second is about requiring history names. The third is about flexible output handlers. The fourth is about overriding parameters. The fifth is about a new XML tag suggestion.

On the right side of the board, there is a "Members" section with a grid of member avatars and a "Board" section with options like "Options", "Add List", and "Filter Cards". Below these is an "Activity" section showing recent actions, such as "Dannon Baker added API: Library Contents to Developer ideas and" and "g2roboto on Feature request: manually hide datasets".

<http://bit.ly/gxyissues>

http://wiki.galaxyproject.org

Galaxy Wiki

FrontPage

Locked History Actions

Galaxy

Galaxy

Community

Conference

OSLO

30 June - 2 July 2013

University of Oslo

Poster abstracts due 3 May

Use Galaxy

Use Main (about) Use Others! Learn Share Search

Communication

Support News Events Twitter Mailing Lists (search)

Deploy Galaxy

Get Galaxy Cloud Admin Tool Config Tool Shed Search

Contribute

Tool Shed Share Issues & Requests Support

Galaxy Project

Home About Community Big Picture

Wiki

Help All Pages

Galaxy

Galaxy is an open, web-based platform for *accessible, reproducible, and transparent* computational biomedical research.

- Accessible:** Users without programming experience can easily specify parameters and run tools and workflows.
- Reproducible:** Galaxy captures information so that any user can repeat and understand a complete computational analysis.
- Transparent:** Users share and publish analyses via the web and create Pages, interactive, web-based documents that describe a complete analysis.

This is the Galaxy Community Wiki. It describes all things Galaxy.

Use Galaxy

Galaxy's [public service web site](#) makes analysis tools, genomic data, tutorial demonstrations, persistent workspaces, and publication services available to any scientist. Extensive [user documentation](#) (applicable to any [public](#) or local Galaxy instance) is available on [this wiki](#) and elsewhere.

Deploy Galaxy

Galaxy is open source for all organizations. Local Galaxy servers can be set up by [downloading and customizing](#) the Galaxy application.

- Admin
- Cloud

Community & Project

Galaxy has a large and active user community and many ways to [Get Involved](#).

- Community
- News
- Events
- Support
- Galaxy Project

Contribute

- Users:** [Share](#) your histories, workflows, visualizations, data libraries, and [Galaxy Pages](#), enabling others to use and learn from them.
- Deployers and Developers:** Contribute tool definitions to the Galaxy [Tool Shed](#) (making it easy for others to use those tools on their installations), and code to the core release.
- Everyone:** [Get Involved!](#)

usegalaxy.org

getgalaxy.org

Events

News

Galaxy Wiki

Events

Locked History Actions

Galaxy Event Horizon

Events with Galaxy-related content are listed here.

Also see the [Galaxy Events Google Calendar](#) for a listing of events and deadlines that are relevant to the Galaxy Community. This is also available as an [RSS feed](#).

If you know of any event that should be added to this page and/or to the Galaxy Event Calendar, please add it here or send it to outreach@galaxyproject.org.

Contents

1. Upcoming Events


2. Other Calendars


3. Past Events


1. 2013


2. Archive


Upcoming Events

 Galaxy Workshop Tour

 GLBIO

 Galaxy Admins

 WINGS

 Galaxy Community Conference 2013

Date	Topic/Event	Venue/Location	Contact
April 29 - May 2	Introduction to Galaxy Workshops 2013 Galaxy Workshop Tour	Washington University in St. Louis	Dave Clements
		Saint Louis University	
		University of Missouri Columbia	
May 14-16	Tutorial: <i>Exploring and Enabling Biomedical Data Analysis with Galaxy</i>	Great Lakes Bioinformatics Conference (GLBIO) 2013, Pittsburgh, Pennsylvania, United States	Anton Nekrutenko
May 15	GalaxyAdmins May 2013 Meetup	GalaxyAdmins web meetup	Srinivas Maddhi, Dave Clements
May 16-17	Galaxy Workflows for Bioinformatics Analysis, and Workshop 1A – Galaxy Workflows for Bioinformatics Analysis	Workshop in Next-Generation Sequence Analysis and Metabolomics (WINGS), UNC-Charlotte, North Carolina, United States	James Taylor
May 21 May 29	Initiation à l'utilisation de Galaxy Les deux ateliers sont maintenant complets		Sandra Dérozier, Valentin Loux, Véronique Martin <veronique.martin AT jouy DOT inra DOT fr>
May 22 May 30	Analyse de données issues de séquenceurs nouvelle génération sous Galaxy Les deux ateliers sont maintenant complets	Cycle "Bioinformatique par la pratique" 2013, INRA Jouy-en-Josas, France	Jean-François Gibrat, Valentin Loux, Véronique Martin <veronique.martin AT jouy DOT inra DOT fr>
May 24 June 19	Introduction to Galaxy	UC Davis Bioinformatics Core Davis, California, United States	Nikhil Joshi <najoshi AT ucdavis DOT edu>
	A Genomics Virtual Lab for Cancer Research		Dominique Gorse
	Next-Gen Sequencing		

Galaxy Wiki

News

Locked History Actions

News

Announcements of interest to the Galaxy Community. These can include items from the Galaxy Team or the Galaxy community and can address anything that is of wide interest to the community.

The Galaxy News is also available as an [RSS feed](#).

See [Add a News Item](#) below for how to get an item on this page, and the RSS feed. Older news items are available in the [Galaxy News Archive](#).

See also

- Galaxy News Briefs
- Galaxy Updates
- Galaxy on Twitter
- Events
- Learn
- Support
- About the Galaxy Project

News Items

Environmental Metabolomics + Galaxy

A new UK-China collaboration in environmental metabolomics between the University of Birmingham, BGI and GigaScience has received funding from the UK's Natural Environment Research Council (NERC).

The first metabolomics project will send a developer from the University of Birmingham's School of Biosciences, to Hong Kong to work with GigaScience personnel on extending Galaxy for use in metabolomics data analyses.

"Metabolomics involves the detection and quantification of small molecules (metabolites) in living organisms and can provide an indication of their cellular condition and health. The toxicological responses of organisms to pollutants can be studied using environmental metabolomics, enabling researchers to discover diagnostic markers for monitoring and risk assessment of our environment. Research at Birmingham focuses extensively on the metabolic responses of the freshwater model organism, *Daphnia*, to both pollutants and engineered nanomaterials."

See the [official announcement](#) for more details.


Peter Li
[GigaScience](#)

Posted to the [Galaxy News](#) on 2013-04-22

Galaxy @ ASMS 2013

Galaxy will have a significant presence at the 61st ASMS Conference on Mass Spectrometry and Allied Topics being held in Minneapolis, Minnesota, June 9-13. Galaxy related content includes the [Galaxy Framework as a Solution for MS-based Informatics](#) workshop and at least 9 posters either directly about or using Galaxy.

If you do research in proteomics than please consider attending.

 Galaxy Community Conference 2013

Poster abstracts due 3 May

Use Galaxy

Use Main (about)
Use Others! • Learn
Share • Search

Communication

Support • News
Events • Twitter
Mailing Lists (search)

Deploy Galaxy

Get Galaxy • Cloud Admin • Tool Config
Tool Shed • Search

Contribute


Tool Shed • Share
Issues & Requests
Support


Galaxy Project

Home • About
Community
Big Picture

Wiki

Help • All Pages
Recent Changes
Search • Create Page

 Galaxy web search

 Galaxy is hiring

galaxyproject.org/GCC2013



STARTING

@

\$125

galaxyproject.org/GCC2013



Poster abstracts due **May 3**



STARTING

@

\$125

The Galaxy Team



Enis Afgan



Dannon Baker



Dan Blankenberg



Dave Bouvier



Dave Clements



Nate Coraor



Carl Eberhard



Dorine Francheteau



Jeremy Goecks



Sam Guerler



Jen Jackson



Greg von Kuster



Ross Lazarus



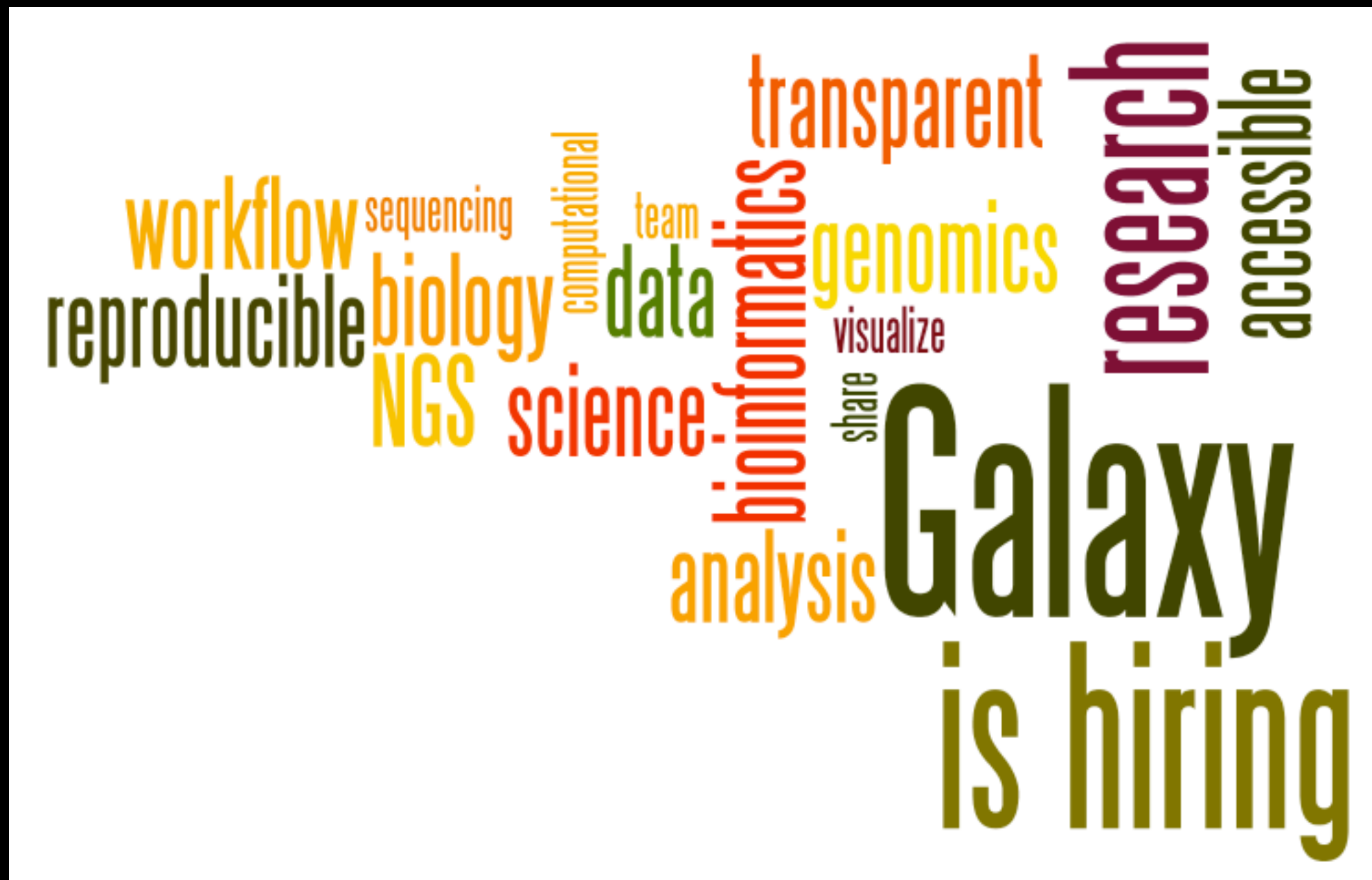
Anton Nekrutenko



James Taylor

<http://wiki.galaxyproject.org/GalaxyTeam>

Galaxy is hiring post-docs and software engineers
at both Emory and Penn State.



Please help.

<http://wiki.galaxyproject.org/GalaxyIsHiring>

Agenda

- 9:00 Welcome
- 9:20 Basic Analysis with Galaxy
- 10:30 Basic Analysis into Reusable Workflows
- 11:00 RNA-Seq Example Part I
- 11:30 Lunch (on your own)
- 1:00 RNA-Seq Example Part II
- 1:30 Galaxy Project Overview
- 2:00 **RNA-Seq Example Part III**
- 2:30 Sharing, Publishing and Reproducibility
- 2:50 Break
- 3:10 ChIP-Seq Example
- 5:00 Done

RNA-seq Exercise: A Plan

- Map the reads to the human reference using Tophat
- Run Cufflinks on Tophat output to assemble reads into transcripts
 - *Imagine pages and pages of discussion on the intricacies and pitfalls of RNA-seq transcript prediction here.*

<http://bit.ly/11Qvnwh>: Lance Parsons' RNA-Seq Tutorial

<http://bit.ly/Y7yKzv>: UC Davis (Nik Joshi?) RNA-Seq Tutorial

<http://bit.ly/YfI0E0>: UAB RNA-Seq Tutorial @ GCC2012

RNA-seq Exercise: A Plan

- Map the reads to the human reference using Tophat
- Run Cufflinks on Tophat output to assemble reads into transcripts
- Visualize it

Visualizing Genomics

Supported external browsers

- UCSC
- Ensembl
- GBrowse
- IGB
- IGV

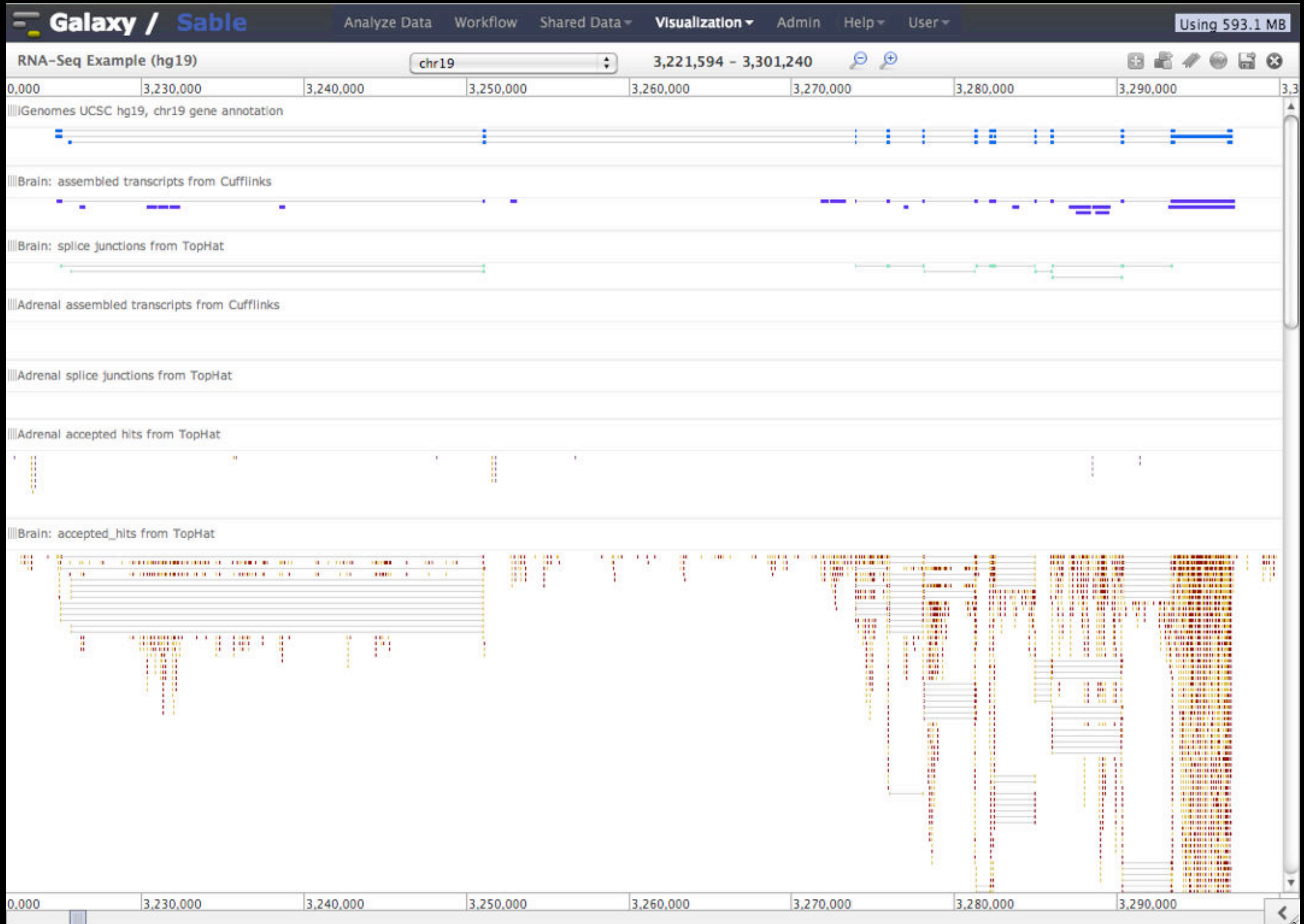
Traditional browser strengths:

- Showing what is nearby
- what else is happening here
- highlighting correlations
- integrating many datasets

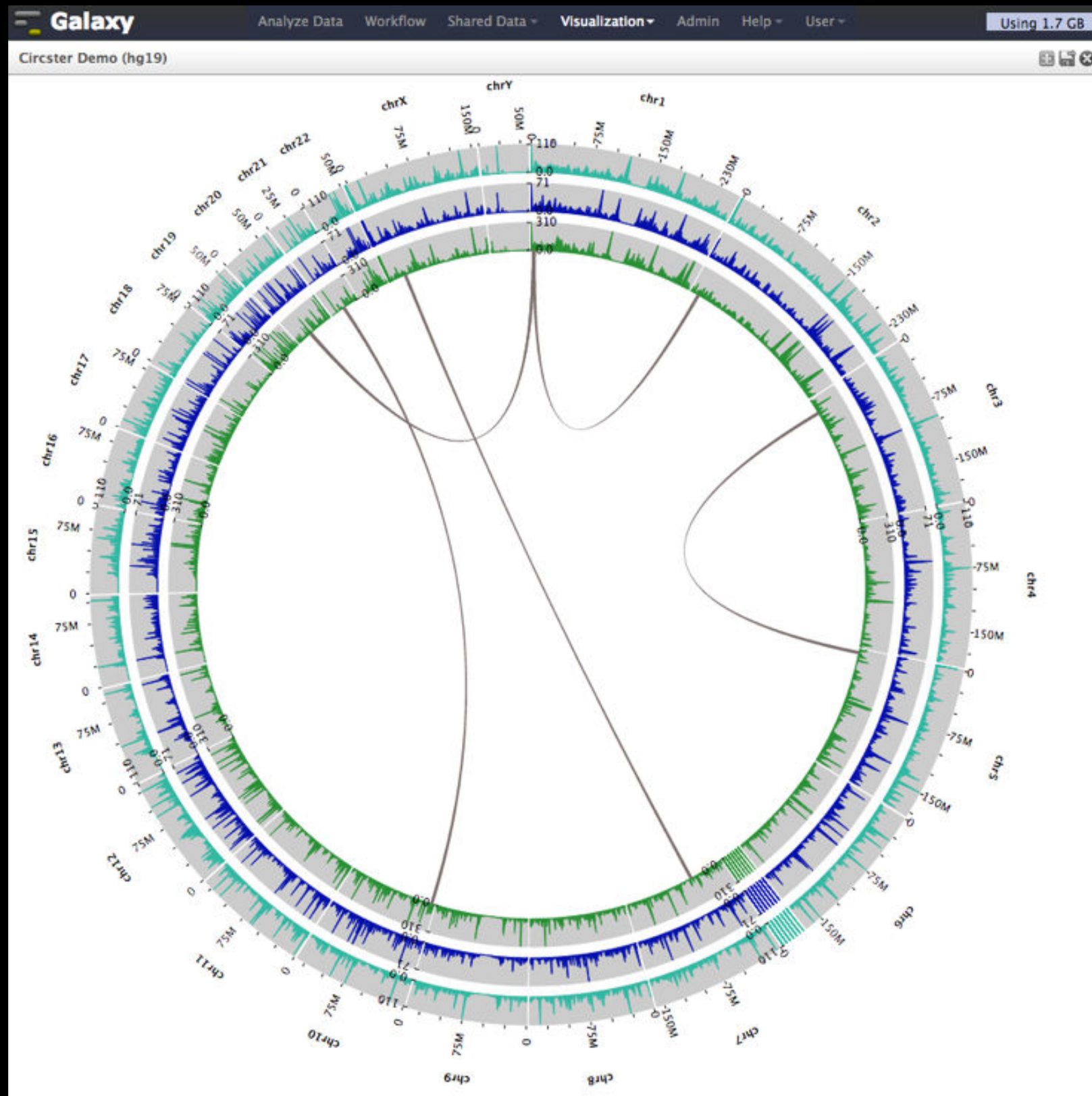
But, *wouldn't it be nice to*

- Use visualization to **evaluate and refine analyses?**
- **Expose** some **basic analyses in visualization** to make it more informative?
- Make that **analyze-visualize-refine** loop seamless and **fast?** That is, integrate the two?
- Use visualization to **learn tools and explore their parameter space?**
- Not be tied to a **predefined reference genome?**

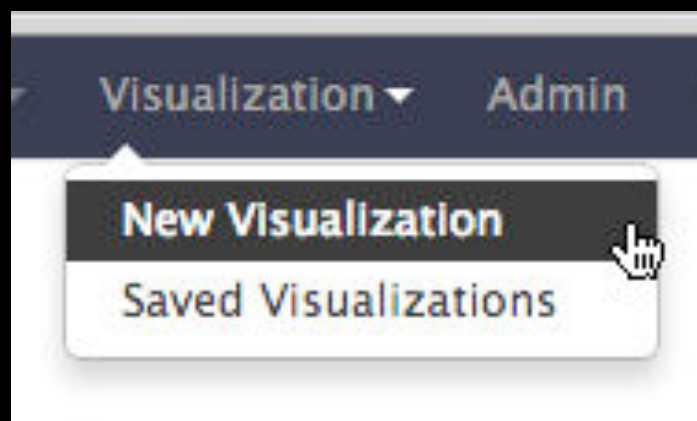
Trackster: Galaxy's embedded track browser



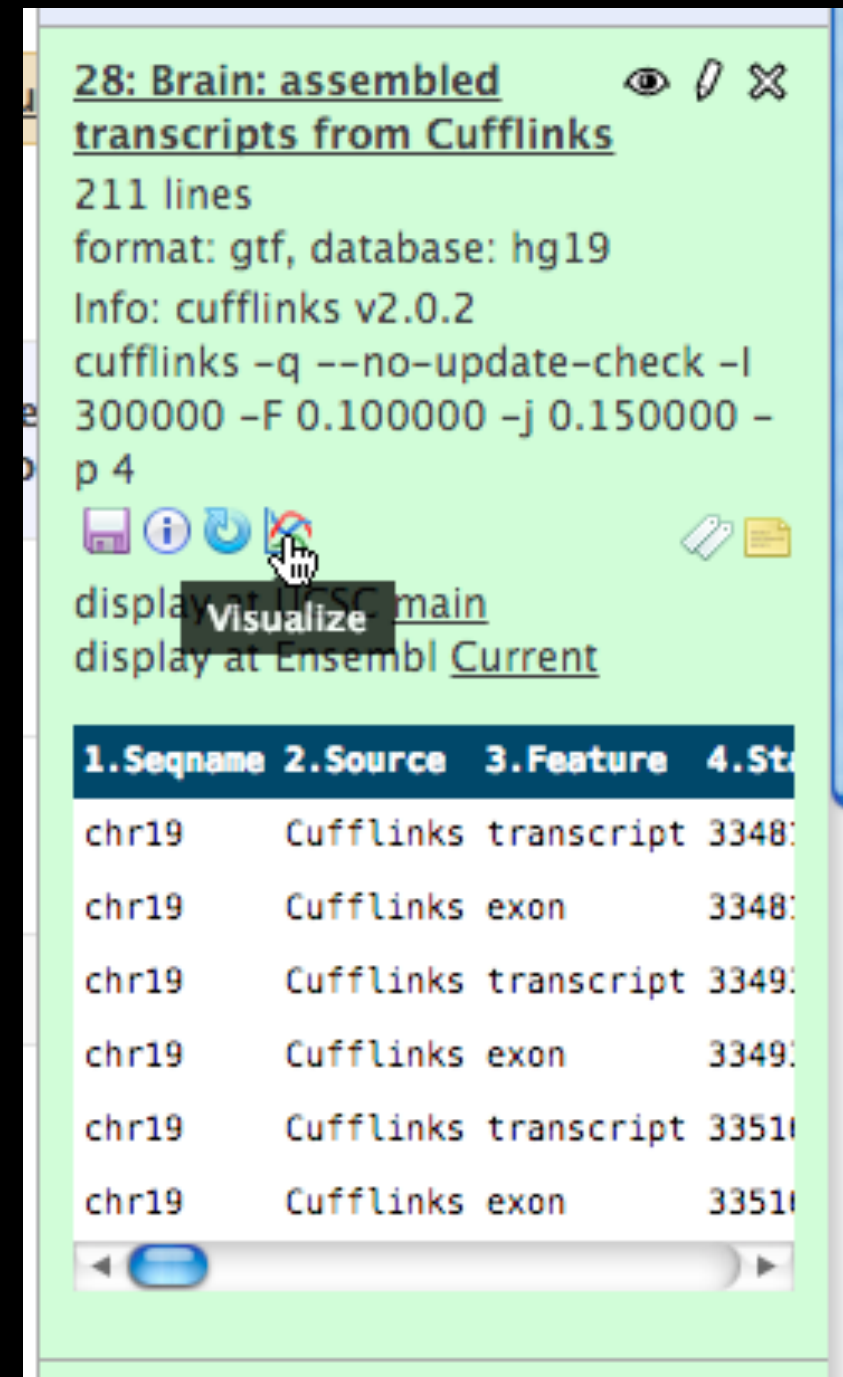
Circster



Create a visualization in Galaxy



or



28: Brain: assembled transcripts from Cufflinks
211 lines
format: gtf, database: hg19
Info: cufflinks v2.0.2
cufflinks -q --no-update-check -l 300000 -F 0.100000 -j 0.150000 -p 4

display at [HSC main](#)
display at Ensembl [Current](#)

1.Seqname 2.Source 3.Feature 4.Start

chr19	Cufflinks	transcript	33480
chr19	Cufflinks	exon	33480
chr19	Cufflinks	transcript	33490
chr19	Cufflinks	exon	33490
chr19	Cufflinks	transcript	33510
chr19	Cufflinks	exon	33510

A screenshot of a Galaxy track visualization. The track is titled '28: Brain: assembled transcripts from Cufflinks' and contains 211 lines of GTF data. The format is 'gtf' and the database is 'hg19'. The track is displayed at the HSC main and Ensembl Current. Below the track is a table with 4 columns: 1.Seqname, 2.Source, 3.Feature, and 4.Start. The table shows 6 rows of data for chr19, including transcripts and exons. A mouse cursor is hovering over the 'Visualize' button.

Isn't it nice to

- To do all those things we talked about?
 - Use visualization to evaluate and refine analyses?
 - Expose some basic analyses in visualization to make it more informative?
 - Make that analyze-visualize-refine loop seamless and fast? That is, integrate the two?
 - Use visualization to learn tools and explore their parameter space?
 - Not be tied to a predefined reference genome?

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More Galaxy Terminology

Share:

Make something available to someone else

Publish:

Make something available to everyone

Galaxy Page:

Analysis documentation within Galaxy; easy to embed any Galaxy object

Sharing & Publishing enables **Reproducibility**

Reproducibility: Everybody talks about it, but ...

Galaxy aims to push the goal of reproducibility from the bench to the bioinformatics realm

All analysis in Galaxy is recorded without any extra effort from the user.

Histories, workflows, visualizations and *pages* can be shared with others or published to the world.

Sharing & Publishing enables **Reproducibility**





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Windshield splatter analysis with the Galaxy metagenomic pipeline

Sergei Kosakovsky Pond^{1,2,6,9}, Samir Wadhawan^{3,6,7},
Francesca Chiaromonte⁴, Guruprasad Ananda^{1,3}, Wen-Yu Chung^{1,3,8},
James Taylor^{1,5,9}, Anton Nekrutenko^{1,3,9} and The Galaxy Team¹

OPEN ACCESS ARTICLE

This Article

Published in Advance October 9, 2009, doi:
10.1101/gr.094508.109

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Current Issue

October 2010, 20 (10)



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Windshield splatter analysis with the Galaxy metagenomic pipeline

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Published in Advance October 9, 2009, doi: 10.1101/gr.094508.109
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Current Issue

October 2010, 20 (10)



Footnotes

[Supplemental material is available online at <http://www.genome.org>. All data and tools described in this manuscript can be downloaded or used directly at <http://galaxyproject.org>. Exact analyses and workflows used in this paper are available at <http://usegalaxy.org/u/aun1/p/windshield-splatter>.]

Windshield splatter analysis with the Galaxy metagenomic pipeline: A live supplement

SERGEI KOSAKOVSKY POND^{1,2,*}, SAMIR WADHAWAN^{3,6*}, FRANCESCA CHIAROMONTE⁴, GURUPRASAD ANANDA^{1,3}, WEN-YU CHUNG^{1,3,7}, JAMES TAYLOR^{1,5}, ANTON NEKRUTENKO^{1,3} and THE GALAXY TEAM^{1*}

Correspondence should addressed to [SKP](#), [JT](#), or [AN](#).

How to use this document


This document is a live copy of supplementary materials for [the manuscript](#). It provides access to the **exact** analyses and workflows discussed in the paper, so you can play with them by re-running, changing parameters, or even applying them to your own data. Specifically, we provide the two histories and one workflow found below. You can view these items by clicking on their name to expand them. You can also import these items into your Galaxy workspace and start using them; click on the green plus to import an item. To import workflows you must [create a Galaxy account](#) (unless you already have one) – a hassle-free procedure where you are only asked for a username and password.

This is the Galaxy history detailing the comparison of our pipeline to MEGAN:




[Galaxy History | Galaxy vs MEGAN](#)


Comparison of Galaxy vs. MEGAN pipeline.

This is the Galaxy history showing a generic analysis of metagenomic data. (This corresponds to the "A complete metagenomic pipeline" section of the manuscript and **Figure 3A**):


[Galaxy History | metagenomic analysis](#)

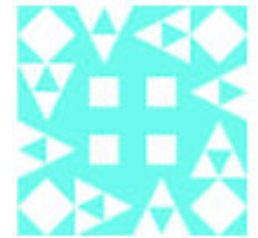

This is the Galaxy workflow for generic analysis of metagenomic data. (This corresponds to the "A complete metagenomic pipeline" section of the manuscript and **Figure 3B**):


[Galaxy Workflow | metagenomic analysis](#)


Generic workflow for performing a metagenomic analysis on NGS data.

Accessing the Data

Windshield Splatter datasets analyzed in this manuscript can be accessed through this [Galaxy Library](#). From there, they can be analyzed through Galaxy using the shared workflows or downloaded.



Author

aun1

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Community
 (6 ratings, 5.0 average)



Tags

Community:

paper galaxy
 megan

<http://usegalaxy.org/u/aun1/p/windshield-splatter>

Sharing for Galaxy Administrators Too

Data Libraries

Make data easy to find

Genome Builds

Care about a particular subset of life?

Galaxy Tool Shed

Wrapping tools and datatypes

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- 5:00 Done

ChIP-Seq Exercise

- Identify zinc-finger CTCF transcription factor tags in mouse
- Example and data from
 - Hillman-Jackson, *et al.*, “Using Galaxy to Perform Large-Scale Interactive Data Analyses” *Curr. Protoc. Bioinform.* 38:10.5.1-10.5.47;
 - ENCODE transcription factor binding experiment: <http://bit.ly/QmD6Nk>. Raw original data generated & analyzed at Michael Snyder’s lab, Stanford University, and Sherman Weissman’s Lab, Yale University.
- We’ll use build **mm9** and datasets that have been prescreened to mostly map to **chr19**
- All datasets are FASTQ

ChIP-Seq Exercise: A Plan

- Get input datasets; control and tags
- Groom the datasets into FASTQSanger format
- Look at quality
- Trim as we see fit
- Map the reads to genome using Bowtie
- Call peaks with MACS (Model-based Analysis of ChIP-seq)

ChIP-Seq Exercise: A Plan

- Get input datasets; control and tags

Shared Data →

Data Libraries →

ChIP-Seq basic datasets

ChIP-Seq Exercise: A Plan

- Get input datasets; control and tags
- Groom the datasets into FASTQSanger format
- NGS: QC and manipulation → **FASTQ Groomer**
 - Input FASTQ quality scores type: **Illumina 1.3-1.7**
 - Run on both datasets

ChIP-Seq Exercise: A Plan

- Get input datasets; control and tags
- Groom the datasets into FASTQSanger format
- Look at quality:
 - NGS QC and Manipulation → FastQC
 - Or one of the other two options we covered earlier

ChIP-Seq Exercise: A Plan

- ...
- Look at quality
- Trim as we see fit:
 - Use one or more of the options described earlier.

Read length is only used for building model to predict fragment length. So if you set fragment size by yourself, it really doesn't matter how long each read is. Also, in MACS models, only 5' ends of each read (only talking about single end sequencing here), where ultrasound or enzymes cut DNA, are informative, for both fragment size prediction and peak calling. So you can still try to let MACS predict fragment size by setting a fixed read length. I think the current cross-correlation way in MACS v2 can give a more stable result than the previous way in MACS v1 just measuring distance between plus and minus read pileup summits.

Tao Liu

https://groups.google.com/forum/?fromgroups=#!topic/macs-announcement/A_Rf0eQ_BLU

ChIP-Seq Exercise: A Plan

- Get input datasets; control and tags
- Groom the datasets into FASTQSanger format
- Look at quality
- Trim as we see fit
- Map the reads to genome using Bowtie
 - NGS: Mapping → Bowtie2
 - Library: Single-end
 - Run on both control and tag files
 - Use mm9 as the reference genome

ChIP-Seq Exercise: A Plan

- Get input datasets; control and tags
- Groom the datasets into FASTQSanger format
- Look at quality
- Trim as we see fit
- Map the reads to genome using Bowtie
- Call peaks with **MACS (Model-based Analysis of ChIP-seq)**

ChIP-Seq Exercise: A Plan

- Call peaks with MACS (Model-based Analysis of ChIP-seq)
 - NGS: Peak Calling → **MACS**
 - Set **ChIP-Seq Tag File** and **ChIP-Seq Control File**
 - Set **Effective genome size: 1.87e+9**
 - Set **Tag size to 36 (still correct?)**
 - Set **Select the regions with MFOLD: 32**
 - Set **Parse xls files into distinct interval files**
 - **Save shifted raw tag count at every bp into a wiggle file**
 - **Resolution for saving wiggle files: 1 (or 10?)**

That's a lot of knobs to set. Get used to it.

Using MACS to Identify Peaks from ChIP-Seq Data

Jianxing Feng,¹ Tao Liu,² and Yong Zhang¹

¹School of Life Sciences and Technology, Tongji University, Shanghai, China

²Department of Biostatistics and Computational Biology, Dana-Farber Cancer Institute and Harvard School of Public Health, Boston, Massachusetts

ABSTRACT

Model-based
Shirley Li
karyotes, c
sites and
control sa

information on how to use MACS to identify either the binding sites of a transcription factor or the enriched regions of a histone modification with broad peaks. Furthermore, the basic ideas for the MACS algorithm and its appropriate usage are discussed. *Curr. Protoc. Bioinform.* 34:2.14.1-2.14.14. © 2011 by John Wiley & Sons, Inc.

Keywords: MACS • ChIP-Seq • peak-calling • cistrome • epigenome

types of histone modifications, the distribution of reads obeys a continuous property, as the epigenetic status of nearby nucleosomes tends to be similar, usually resulting in quite broad peaks. With proper parameter settings, MACS performs well to detect histone-modification-enriched regions. Similarly, MACS can also be applied in affinity enrichment-based DNA methylation studies, such as MeDIP-Seq data.

Know what you are doing

⚠ There is no such thing (yet) as an automated gearshift in short read mapping. It is all like stick-shift driving in San Francisco. In other words = running this tool with default parameters will probably not give you meaningful results. A way to deal with this is to **understand** the parameters by carefully reading the documentation and experimenting. Fortunately, Galaxy makes experimenting easy.

Identifying ChIP-seq enrichment using MACS

Jianxing Feng^{1,3}, Tao Liu^{2,3}, Bo Qin¹, Yong Zhang¹ & Xiaole Shirley Liu²

¹Department of Bioinformatics, School of Life Sciences and Technology, Tongji University, Shanghai, China. ²Department of Biostatistics and Computational Biology, Harvard School of Public Health, Dana-Farber Cancer Institute, Boston, Massachusetts, USA. ³These authors contributed equally to this work. Correspondence should be addressed to Y.Z. (yzhang@tongji.edu.cn) and X.S.L. (xslu@jimmy.harvard.edu).

Published online 30 August 2012; doi:10.1038/nprot.2012.101

Open Access

Method

Model-based Analysis of ChIP-Seq (MACS)

Yong Zhang^{✉*}, Tao Liu^{✉*}, Clifford A Meyer^{*}, Jérôme Eeckhoutte[†], David S Johnson[‡], Bradley E Bernstein^{§¶}, Chad Nusbaum[¶], Richard M Myers[¥], Myles Brown[†], Wei Li[#] and X Shirley Liu^{*}

Addresses: ^{*}Department of Biostatistics and Computational Biology, Dana-Farber Cancer Institute and Harvard School of Public Health, 44 Binney Street, Boston, MA 02115, USA. [†]Division of Molecular and Cellular Oncology, Department of Medical Oncology, Dana-Farber Cancer Institute and Department of Medicine, Brigham and Women's Hospital and Harvard Medical School, 44 Binney Street, Boston, MA 02115, USA. [‡]Gene Security Network, Inc., 2686 Middlefield Road, Redwood City, CA 94063, USA. [§]Molecular Pathology Unit and Center for Cancer Research, Massachusetts General Hospital and Department of Pathology, Harvard Medical School, 13th Street, Charlestown, MA 02129, USA. [¶]Broad Institute of Harvard and MIT, 7 Cambridge Center, Cambridge, MA, 02142, USA. [¥]Department of Genetics, Stanford University Medical Center, Stanford, CA 94305, USA. [#]Division of Biostatistics, Dan L Duncan Cancer Center, Department of Molecular and Cellular Biology, Baylor College of Medicine, One Baylor Plaza, Houston, TX 77030, USA.

✉ These authors contributed equally to this work.

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Published: 17 September 2008

Genome Biology 2008, 9:R137 (doi:10.1186/gb-2008-9-9-r137)

The electronic version of this article is the complete one and can be found online at <http://genomebiology.com/2008/9/9/R137>

Received: 4 August 2008

Revised: 3 September 2008

Accepted: 17 September 2008

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- 3:10 ChIP-Seq Example
- 5:00 **Done, almost**

Feedback

<http://bit.ly/20130430Gxy>

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Maureen Donlin
Kristi Holmes
Bob Engeszer

The Galaxy Team
You!

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Saint Louis University

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Penn State University Emory University

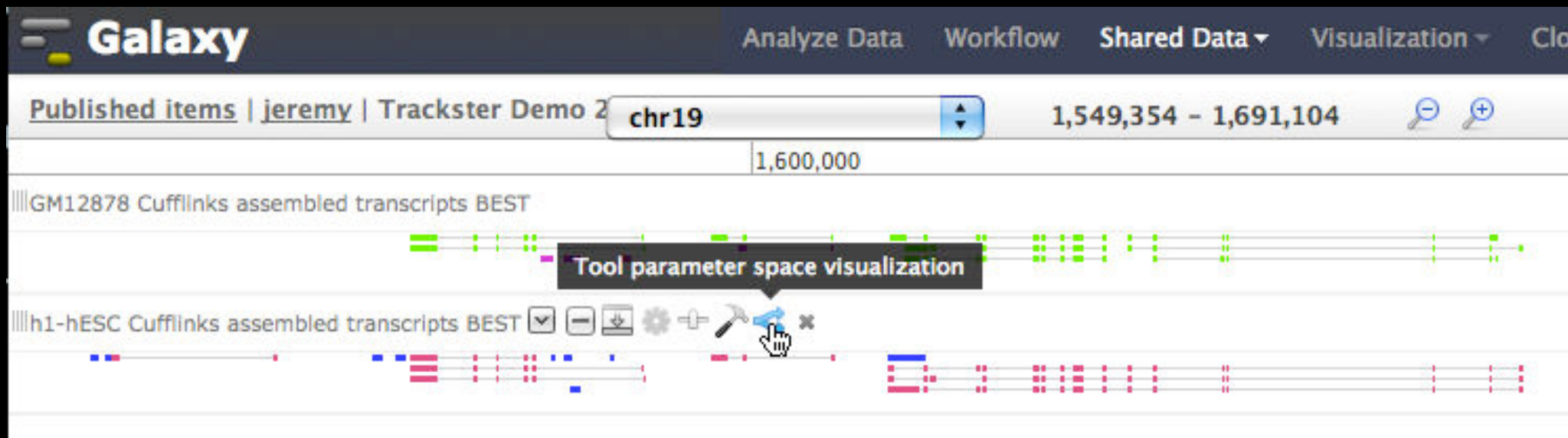
<http://bit.ly/20130430Gxy>



Thanks

<http://bit.ly/20130430Gxy>

Exploring Parameter Space with Trackster



Galaxy interface showing the Cufflinks (version 0.0.5) tool configuration and a parameter space visualization tree.

Cufflinks (version 0.0.5)

- ☐ Max Intron Length: 200000 - 400000 samples: 3
- ☐ Min Isoform Fraction: 0.1 - 0.2 samples: 3
- ☒ Pre mRNA Fraction: 0.15
- ☐ Perform quartile normalization: No, Yes
- ☒ Use multi-read correct: No

Getting Started

1. Create a parameter tree by using the icons next to the tool's parameter names to add or remove parameters.
2. Adjust the tree by using parameter inputs to select min, max, and number of samples
3. Run the tool with different settings by clicking on tree nodes

Parameter Space Visualization Tree:

- Root
 - Max Intron Length
 - 200000
 - Perform quartile normalization
 - No
 - Min Isoform Fraction
 - 0.1
 - 0.15
 - 0.2
 - Yes
 - Min Isoform Fraction
 - 0.1
 - 0.15
 - 0.2
 - 300000 (selected)
 - 400000
 - Perform quartile normalization
 - No
 - Min Isoform Fraction
 - 0.1
 - 0.15
 - 0.2
 - Yes
 - Min Isoform Fraction
 - 0.1
 - 0.15
 - 0.2

Galaxy CloudMan

<http://usegalaxy.org/cloud>

- Start with a **fully configured and populated** (tools and data) Galaxy instance.
- Allows you to scale up and down your compute assets as needed.
- Someone else manages the data center.
- **We are using this today.**



- **You will set up an instance now**

<http://aws.amazon.com/education>

Could do this step by step, but ...

<http://bit.ly/GXYAWSGetStarted>

Galaxy Wiki

CloudMan/AWS/GettingStarted

Login | Search:

Getting Started with Galaxy CloudMan

This page provides a step-by-step instructions on how to start your own instance of Galaxy on [Amazon Web Services \(AWS\) Elastic Compute Cloud \(EC2\)](#). More general information and instructions about Galaxy CloudMan (GC) can be found [here](#).

Contents

1. [Step 1: One Time Amazon Setup](#)
2. [Step 2: Starting a Master Instance](#)
3. [Step 3: Galaxy CloudMan Web Interface](#)
4. [Step 4: Use Galaxy as you normally would](#)
5. [Step 5: Shutting Down](#)

AWS

Get Started

Capacity Planning

AMIs

↑ CloudMan

Step 1: One Time Amazon Setup

1. Because AWS services implement pay-as-you-go access model for compute resources, it is necessary for every user of the service to [register with Amazon](#). You will need a credit card to register. (You can apply for a [AWS Education Grant](#) after you register).
2. Once your account has been approved by Amazon (note that this may take up to one business day), [log into the EC2 AWS Management Console](#) and set your AWS Region to *US East (Virginia)*. This is the only region Galaxy CloudMan is fully supported in at this time (see [screenshot 1.2](#)).
3. Click **Network & Security** → **Key Pairs** or **My Resources** → *n* **Key Pairs** (see [screenshot 1.3](#) - if it does not look like this, then try using the Chrome browser) and then click **Create Key Pair**. Enter a memorable name for the key pair, e.g., GalaxyCloud and click **Create**.
4. *Save your private key!* The previous step creates the key pair and downloads a copy to your machine with the name *MemorableName.pem*. Save this file and protect it like you would your password. The key pair can be used to access started instances from

Step 1 Screenshots



1.2. Set region



Instant CloudMan

<http://usegalaxy.org/cloudlaunch>

The image shows two overlapping screenshots of the Galaxy web interface. The top screenshot displays the main Galaxy dashboard with a 'Cloud' dropdown menu open, showing the option 'New Cloud Cluster'. The bottom screenshot shows the 'Launch a Galaxy Cloud Instance' form, which includes fields for Cluster Name, Password, Key ID, Secret Key, and Instance Share String (optional). The Instance Type is set to 'Large'. A 'Submit' button is at the bottom of the form.

Galaxy Analyze Data Workflow Shared Data Visualization Cloud Help User Using 0%

Tools

search tools

Get Data

- [Upload File](#) from your computer
- [UCSC Main](#) table browser
- [UCSC Archaea](#) table browser
- [BX main](#) browser
- [EBI SRA](#) ENA SRA
- [BioMart](#) Central server
- [GrameneMart](#) Central server
- [Flymine](#) server
- [modENCODE fly](#) server
- [modENCODE modMine](#) server

Managing Data
Store, Manage, and Share data with Libraries
An in-depth tutorial

0 bytes

Your history is empty. Click 'Get Data' on the left pane to start

Galaxy Analyze Data Workflow Shared Data Visualization Cloud Help User Using 0%

Launch a Galaxy Cloud Instance

Cluster Name

Password

Key ID

Secret Key

Instance Share String (optional)

Instance Type
Large

Requesting the instance may take a moment, please be patient. Do not refresh your browser or navigate away from the page

Instant CloudMan

AWS Credentials

[**http://bit.ly/**](http://bit.ly/)