

Using Galaxy to provide a NGS Analysis Platform

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Friedrich Miescher Institute

- part of the Novartis Research Foundation
- affiliated institute of Basel University
- member of Swiss Institute of Bioinformatics

316 employees

(incl. 96 PhD students, 95 Post Docs)

Epigenetics

(8 research groups)

Growth Control

(7 research groups)

Neurobiology

(8 research groups)

Technology Platforms

Computational Biology – Cell Sorting – Imaging and Microscopy –
Functional Genomics – Histology – Mass Spectrometry – Protein Structure



Swiss Institute of
Bioinformatics



NOVARTIS



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working with NGS data is *fascinating* because

- there is a different instrument on the market every few months
- Scientists come up with new kind experiments
- new algorithms to deal with NGS data are developed continuously

working with NGS data is *difficult* because

*people with different background/training
are interested in using NGS*

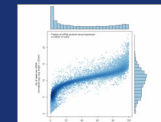
the “average” lab scientist is looking for the red button to press

bizarre output from
the sequencer



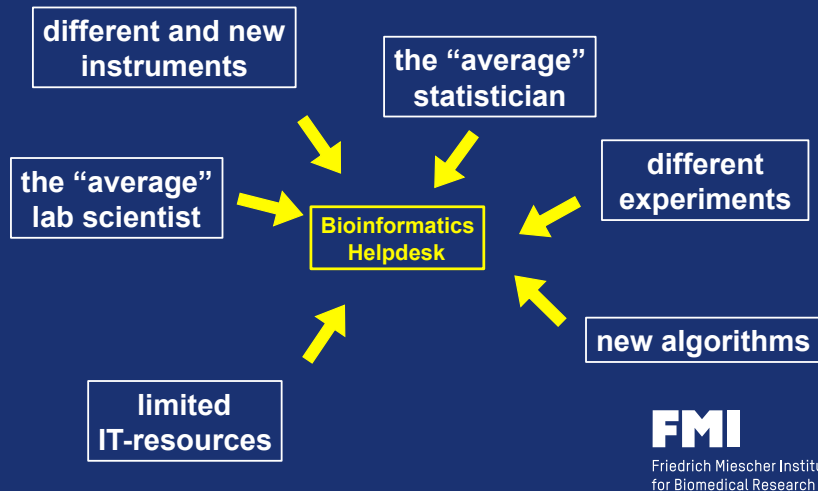
publication in *Nature*

the “average” statistician is creating wonderful blots.....

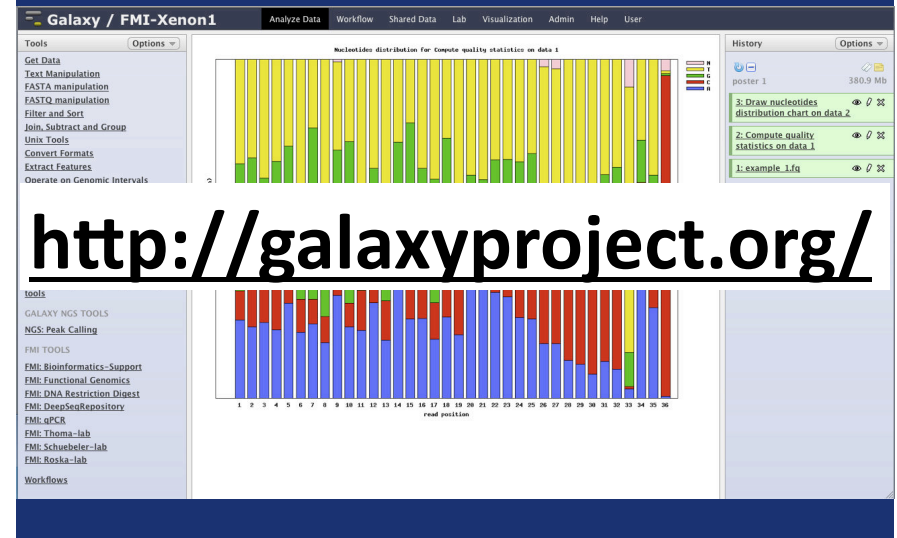


...nobody understands

and the Bioinformatics Helpdesk is caught
in the middle....



the solution:



<http://galaxyproject.org/>

<http://galaxyproject.org/>

Galaxy

“Galaxy is an open, web-based platform for data intensive biomedical research. Whether on the free public server or your own instance, you can perform, reproduce, and share complete analyses.”

The Galaxy Team is a part of BX at Penn State, and the Biology and Mathematics and Computer Science departments at Emory University.

The Galaxy Project is supported in part by NSF, NHGRI, The Huck Institutes of the Life Sciences, The Institute for CyberScience at Penn State, and Emory University.

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<http://galaxyproject.org/>

Galaxy

....and I am NOT part of the Galaxy Team!

I am just a member of the worldwide community of many Galaxy users, adopters, developers, evangelists, etc.

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what is Galaxy?



- provides a GUI to Bioinformatics tools
- manages/stores your (raw) data and results
- allows you to create workflows
- allows sharing and reproducing your analysis

public Galaxy instance:

<http://usegalaxy.org>

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why are we using Galaxy



- open source
(<http://wiki.g2.bx.psu.edu/Admin/License>)
- we can modify the tools
- we can add our own tools
- it is part of a wider community:
“GenomeSpace”, “GMOD”
- it is flexible and simple to install

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it is really simple to install



requirements:

- Python (2.5 or 2.6)
- Mercurial

just 3 commands:

- hg clone <https://bitbucket.org/galaxy/galaxy-dist/>
- cd galaxy_dist
- sh run.sh

...and it is ready (on linux and Mac) at:

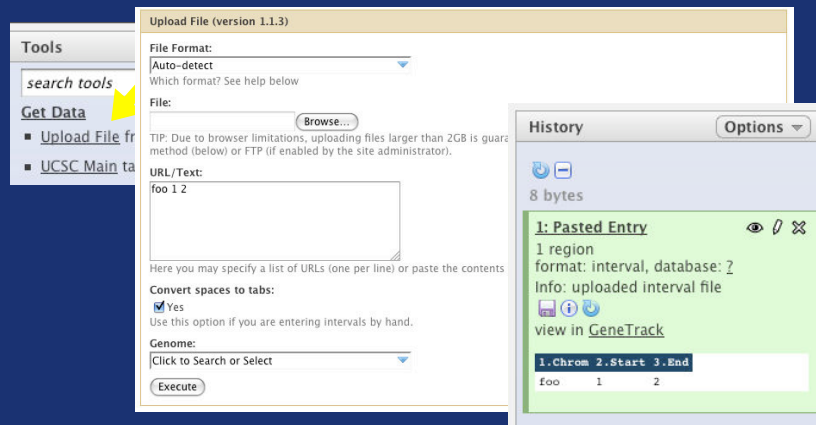
<http://localhost:8080>

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The screenshot shows the Galaxy web interface. On the left is a 'Tools' panel with a search bar and a list of tool categories including 'Get Data', 'Text Manipulation', 'Statistics', and 'NGS'. The central area displays a workflow diagram titled 'WWFSMD? grow noody appendages...'. On the right is a 'History' panel showing a list of data objects. Overlaid on the screenshot are the words 'Tools', 'GUI', and 'Display' in large, bold, black text.

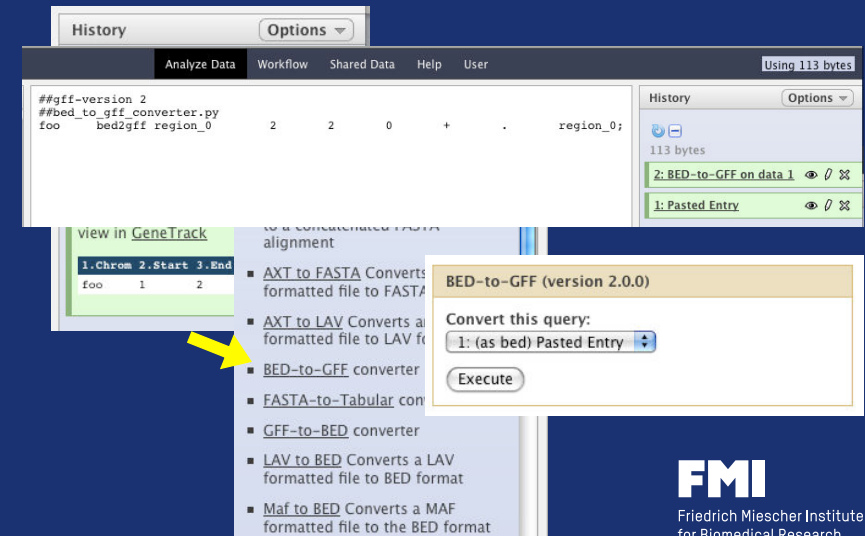
how does it work



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how does it work



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what kind of tools do you get



input tools:

- text box / upload file / url
- access to a local file system ("Data Libraries")
- access to UCSC table browser and ensembl biomart

text manipulation tools:

- file conversion
- table calculation
- operation on genomic intervals

wrappers (and GUIs):

EMBOSS, NCBI BLAST+, NGS: QC and manipulation, Picard, NGS: Mapping, NGS: Indel Analysis, NGS: RNA Analysis, SAM Tools, GATK Tools, NGS: Peak Calling, and much more...

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what needs to be added



EMBOSS, NCBI BLAST+, bowtie, BWA, macs, cufflinks, samtools, etc
plus the corresponding databases, indices, etc

yes, this is annoying.....

but, provides a lot of flexibility.....

and if you don't want to use eg BLAST, remove it from the list and don't care about the binary.

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do you really need all the tools



NGS: QC and manipulation

- FASTQC: FASTQ/SAM/BAM
 - Fastqc: Fastqc QC using FastQC from Babraham
- ILLUMINA FASTQ
 - FASTQ Groomer convert between various FASTQ quality formats
- FASTQ splitter on joined paired end reads
- FASTQ joiner on paired end reads
- FASTQ Summary Statistics by column
 - ROCHE-454 DATA
- Build base quality distribution
- Select high quality segments
- Combine FASTA and QUAL into FASTQ
- AB-SOLID DATA
 - Convert SOLID output to fastq
- Compute quality statistics for SOLID data
- Draw quality score boxplot for SOLID data

FASTX-TOOLKIT FOR FASTQ DATA

- Quality format converter (ASCII-Numeric)
- Compute quality statistics
- Draw quality score boxplot
- Draw nucleotides distribution chart
- FASTQ to FASTA converter
- Filter by quality
- Remove sequencing artifacts
- Barcode Splitter
- Clip adapter sequences
- Collapse sequences
- Rename sequences
- Reverse-Complement
- Trim sequences

GENERIC FASTQ MANIPULATION

- Filter FASTQ reads by quality score and length
- FASTQ Trimmer by column
- FASTQ Quality Trimmer by sliding window
- FASTQ Masker by quality score
- FASTQ interlacer on paired end reads
- FASTQ de-interlacer on paired end reads
- Manipulate FASTQ reads on various attributes
- FASTQ to FASTA converter
- FASTQ to Tabular converter
- Tabular to FASTQ converter



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do you really need all the tools



NGS: Mapping

- Lastz map short reads against reference sequence
- Lastz paired reads map short paired reads against reference sequence
- Map with Bowtie for Illumina
- Map with Bowtie for SOLiD
- Map with BWA for Illumina
- Map with BWA for SOLiD
- Map with BFAST
- Megablast compare short reads against htgs, nt, and wgs databases
- Parse blast XML output
- Map with PerM for SOLiD and Illumina
- Re-align with SRMA
- Map with Mosaik

NGS: RNA Analysis RNA-SEQ

- Tophat for Illumina Find splice junctions using RNA-seq data
 - Tophat for SOLiD Find splice junctions using RNA-seq data
 - Cufflinks transcript assembly and FPKM (RPKM) estimates for RNA-Seq data
 - Cuffcompare compare assembled transcripts to a reference annotation and track Cufflinks transcripts across multiple experiments
 - Cuffdiff find significant changes in transcript expression, splicing, and promoter use
- FILTERING
- Filter Combined Transcripts using tracking file

NGS: SAM Tools

- Filter SAM on bitwise flag values
- Convert SAM to interval
- SAM-to-BAM converts SAM format to BAM format
- BAM-to-SAM converts BAM format to SAM format
- Merge BAM Files merges BAM files together
- Generate pileup from BAM dataset
- Filter pileup on coverage and SNPs
- Pileup-to-Interval condenses pileup format into ranges of bases
- flagstat provides simple stats on BAM files
- rmDup remove PCR duplicates



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from the laptop to a production environment

- remove the tools you don't want
- switch from SQLite to PostgreSQL or MySQL
- use a proxy server
- authenticate users externally via Kerberos or LDAP
- use a 'big' server
- use a compute cluster (TORQUE PBS, PBS Pro, Platform LSF, and Sun Grid Engine)



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or use the "cloud"



<http://usegalaxy.org/cloud>
(using "Amazon Elastic Compute Cloud")

Enis Afgan, et al.
Harnessing cloud computing with Galaxy Cloud
Nature Biotechnology 29, 972-974,
Published online 08 November 2011
www.nature.com/nbt/journal/v29/n11/full/nbt.2028.html



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adding your own tools



everything is possible in Galaxy

As long as you can run it on the command line, you can incorporate it into Galaxy.

- add the executable or script (perl, python, bash, R, etc)
- write a tool definition file
- add it to the list of tools

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tool definition file



```
<tool id="bed2gff1" name="BED-to-GFF" version="2.0.0">
<description>converter</description>

<command>bed_to_gff_converter.py $input $out_file1</command>

<inputs>
  <param format="bed" name="input" type="data" label="Convert this"/>
</inputs>

<outputs>
  <data format="gff" name="out_file1" />
</outputs>

<help>
This tool converts data from BED format to GFF format
</help>
</tool>
```

➔ no need to define/design a GUI !

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Galaxy Tool Shed



enables sharing of tools across the Galaxy community.

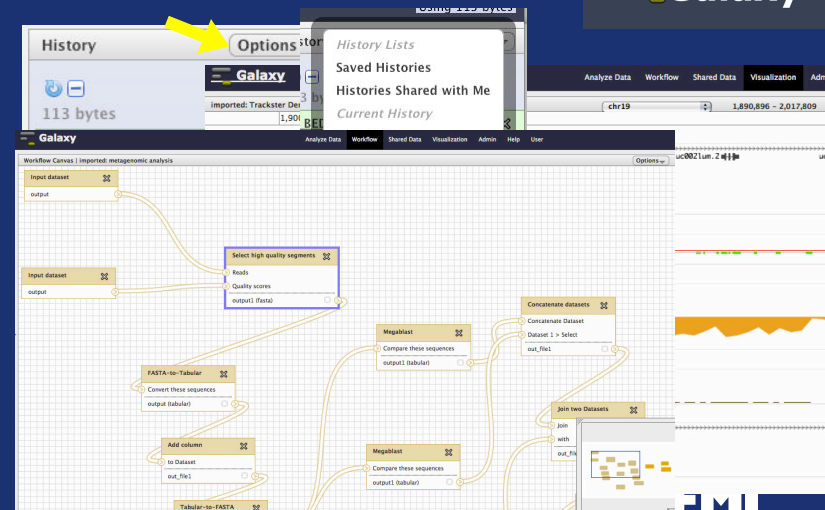
Name	Description	Repositories
Assembly	Tools for working with assemblies	12
Computational chemistry	Tools for use in computational chemistry	2
Convert Formats	Tools for converting data formats	13
Data Source	Tools for retrieving data from external data sources	3
Fasta Manipulation	Tools for manipulating fasta data	17
Genomic Interval Operations	Tools for operating on genomic intervals	0
Graphics	Tools producing images	8
Next Gen Mappers	Tools for the analysis and handling of Next Gen sequencing data	24
Ontology Manipulation	Tools for manipulating ontologies	3
SAM	Tools for manipulating alignments in the SAM format	8
Sequence Analysis	Tools for performing Protein and DNA/RNA analysis	44
SNP Analysis	Tools for single nucleotide polymorphism data such as WGA	4
Statistics	Tools for generating statistics	8
Text Manipulation	Tools for manipulating data	14
Visualization	Tools for visualizing data	9

<http://toolshed.g2.bx.psu.edu/>

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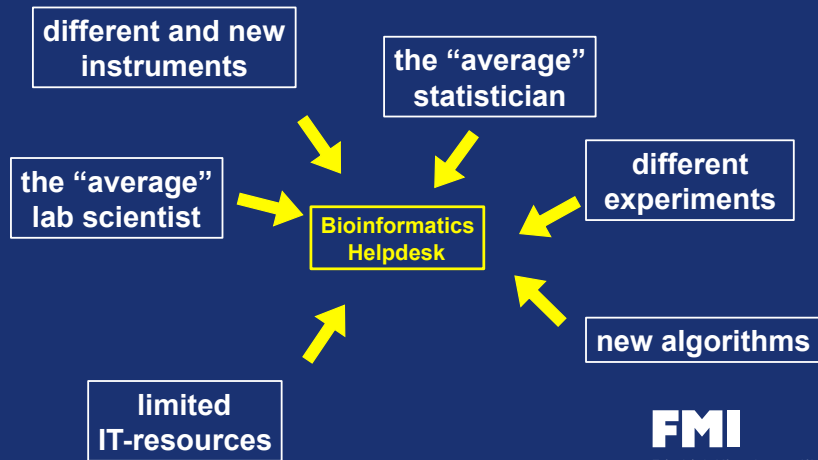
a few more highlights



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what happened to the poor guy sitting at the Bioinformatics Helpdesk....



He is a Galaxy administrator



...and he adds the R script to Galaxy

→ the lab scientist can repeat the analysis
in the 'friendly' web browser and
not on the 'scary' command line

He is a Galaxy administrator



...and he adds the new tools to Galaxy

→ and everybody can test them
without any delay

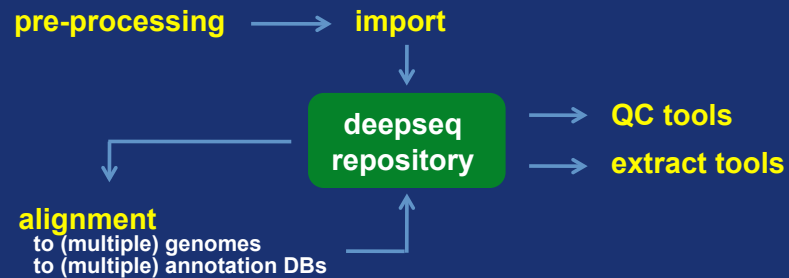
He is a Galaxy administrator



limited
IT-resources

- shared infrastructure
- Galaxy provides 'report' tools

NGS analysis at the FMI



The NGS pipeline at the FMI is



....just a bunch of Perl scripts *(currently)*

➔ which can be easily added to Galaxy

....just a simple file system *(currently)*

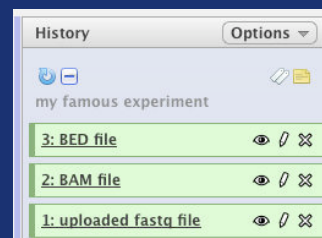
....which cannot be added to Galaxy.
(Galaxy uses its own data directory)

➔ we don't have to, we just have to give
Galaxy access to the directory
(without using "Data Libraries")

a simple NGS workflow



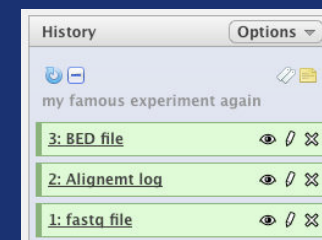
- your famous aligner
- your famous extract tool



a simple NGS workflow

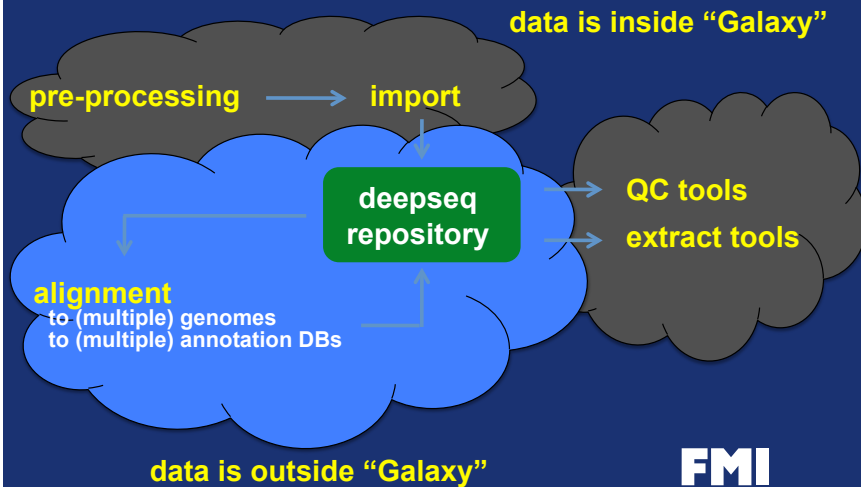


- do you need the result (ie the alignment)
as a new history item?
- does your tool require a Galaxy history item as input?



- the 'famous aligner' has a wrapper
storing the BAM file in the central
NGS repository and creating just
a log file for Galaxy
- your 'famous extract tool' knows
the location of the NGS repository

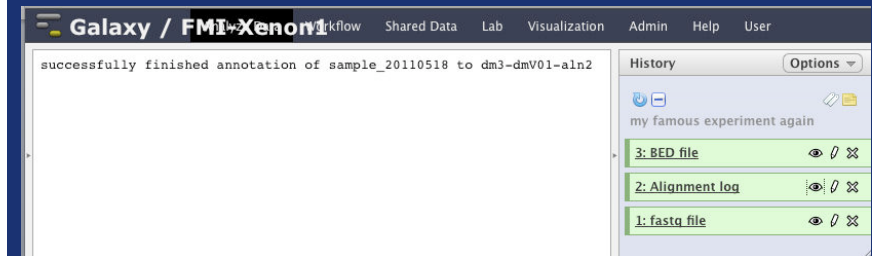
NGS analysis at the FMI



storing data outside of Galaxy



makes it easier to share with non-Galaxy users



makes it easier to share with non-Galaxy users

```
successfully finished annotation of  
sampleId_20110518 to dm3-dmV01-aln2
```

and now the command line geek can do

```
[geek@xenon1 ~]$ extractData.pl -f -s p -m  
100 -i mySampleId_20110518 dm3-dmV01-aln2  
genome | frag2bed.pl -t -q -U - | head -5  
track name='mySampleId_20110518'  
chr2L 10493 10528 sq39319 1 +  
chr2L 10736 10764 sq74484 1 +  
chr2L 11442 11477 sq1340 1 +  
chr2L 13799 13834 sq84955 1 +  
[geek@xenon1 ~]$
```

makes it easier to share with non-Galaxy users

command line

```
extractData.pl -f -s p -m 100 -i  
mySampleId_20110518 dm3-dmV01-aln2 genome |  
frag2bed.pl -t -q -U -
```

Galaxy tool definition file

```
#elif ($summary.mode=="bed")#extractData.pl  
-f $strand $maxhits $signCnts  
$sampleSelect.sampleId $genome-$annot-aln2  
genome | frag2bed.pl -t -q $summary.ucsc -  
> $output
```

and doing the same in Galaxy



Extract data (step 1 of 2)

Sample selection:

Extract data (step 2 of 2)

Strand selection:
Ignore strand (use all)

History

Options

my famous experiment again 17.9 Mb

3: BED file

2: Alignment log

1: fastq file

track	name	mySampleId_20110518'
chr2L	10493	10528
chr2L	10736	10764
chr2L	11442	11477
chr2L	13799	13834
chr2L	13940	13974
chr2L	13948	13979
chr2L	14266	14301
chr2L	14381	14414
chr2L	14612	14645
chr2L	15215	15250
chr2L	18459	18490
chr2L	21264	21295
chr2L	67455	67489
chr2L	72882	72916
chr2L	75216	75251
chr2L	75381	75416
chr2L	75416	75451
chr2L	76053	76088
chr2L	85320	85355
chr2L	101308	101343
chr2L	102620	102655
chr2L	103097	103132
chr2L	103605	103640
chr2L	103769	103802
chr2L	103855	103890

Summary

Mission

running a Bioinformatics Helpdesk

Vision

I don't have to do anything

Strategy



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Acknowledgment



Michael Stadler Lukas Burger
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Anita Lerch
Thomas Übermeier Jan Welker

....and all the people from the "Galaxy"



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a few web sites



<http://galaxyproject.org>

<http://usegalaxy.org>

<http://usegalaxy.org/galaxy101>

<http://usegalaxy.org/cloud>

<http://toolshed.g2.bx.psu.edu/>

<http://wiki.g2.bx.psu.edu/Admin/License>



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