

INTRODUCTION

Proteogenomics and metaproteomics are rapidly emerging fields, based on integrated analysis of multi-omic data. For many studies of human health and disease, combining proteogenomic and metaproteomic analysis offers great promise for new discoveries. Proteogenomics integrates high throughput genomic/transcriptomic sequencing data with MS-based proteomic data, identifying novel, diseaseassociated proteoforms in the human host. Metaproteomics integrates metagenomic data with MS-based proteomic data, identifying disease-associated proteins expressed by microbial communities residing in the host. Informatic workflows for proteogenomics and metaproteomics are inherently complex, and not run in combination. Consequently, neither tandem proteogenomicmetaproteomic analysis, nor the new discoveries it offers, is accessible to most researchers. In response, we describe a solution - the Galaxy framework for bioinformatic workflow development. Galaxy offers many unique and powerful features for multi-omic applications, including the ability to integrate numerous disparate software and processing steps, into automated workflows. These complete workflows can be shared with others, promoting reproducibility and accessibility. Capitalizing on Galaxy's flexibility, we have developed a novel tandem proteogenomic-metaproteomic workflow, presented here.



We are developing Galaxy for proteomics, or Galaxy-P, which is an extension of the Galaxy framework implementing software and tools for MS-based proteomic and multi-omic analysis applications. Unique benefits of Galaxy-P include:

- Flexibility for integrating established and new software tools into customized workflows
- Accessibility for biologists (usegalaxyp.org)
- Share-ability of tools, complete workflows and histories promoting reproducibility, transparency and community evaluation

Galaxy-P Tools



transcriptomic data analysis



modular workflows that can be reused, shared and creatively modified for multiple studies.

METHODS

RAW files from multiple datasets (see below) were generated from Orbitrap XL instrument. The processed peak lists were searched using ProteinPilot [™] version 4.5 (AB Sciex) within Galaxy-P. After optimization and testing, multiple workflows were used in a sequential manner to generate inputs for the subsequent workflow.

• Salivary supernatant was 3D-fractionated with or without ProteoMiner treatment (Bandhakavi et al 2009) was used. 200 RAW files were acquired on LTQ/Orbitrap XL. Both the datasets were searched against the human oral microbiome database (HOMD) or the 3-frame translated human cDNA database using the "Minnesota two-step" method (Jagtap et al 2013)

CUSTOMIZED, REPRODUCIBLE AND ACCESSIBLE TOOLS FOR TANDEM PROTEOGENOMIC-METAPROTEOMIC ANALYSIS

UNIVERSITY OF MINNESOTA Pratik Jagtap¹, James Johnson¹, John Chilton^{1,3}, Trevor Wennblom¹, Bart Gottschalk¹, Sricharan Bandhakavi^{1,2}, Joel Rudney¹, Tim Griffin¹ ¹University of Minnesota, Minneapolis, MN; ²Bio-Rad Laboratories, Hercules, CA; ³Pennsylvania State University, University Park, PA



ks loaded chr12:11,545,849



<u>ESULTS</u>						
iom	ICS					
orm ed)	Distinct, peptide sequences mapped to genome					
	52					
	Normal coding frame Alternative frame Exon Intron or untranslated region					
ne 12 3.1 p12	.3 p12.2					
oing to the dentified ces from erminus) uous fran	PRB2 11,549 kb 11,549 kb 11,540					
	aice					

Novel proteo

peptides (filt

105

luman chromosor

microbial origi

1926

Metaproteomics

Phyla	Genera	Species
12	65	123



Functional analysis of microbial proteins in whole saliva analyzed via the MEGAN software for taxonomic and bioinformatic analysis of large-scale metagenomic or,

REMA	RKS					
passed	flexibility		for			
ding pow	erful w	orkfl	OWS			
ic and	metap	rotec	omic			
tomated [BLASTP					
a means t	o ensu	re hi	gh			
alyses						
V for visualizing novel						
s) and MEGAN or						
al analysis						
eps) can be run with little						
in their c	omplet	enes	S			
		wash	۲)			
arant 11	17070	INCE		/		